

Package ‘gclink’

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Title Gene-Cluster Discovery, Annotation and Visualization

Version 1.1

Description Performs end-to-end analysis of gene clusters—such as photosynthesis, carbon/nitrogen/sulfur cycling, carotenoid, antibiotic, or viral marker genes (e.g., capsid, polymerase, integrase)—from genomes and metagenomes. It parses Basic Local Alignment Search Tool (BLAST) results in tab-delimited format produced by tools like NCBI BLAST+ and Diamond BLASTp, filters Open Reading Frames (ORFs) by length, detects contiguous clusters of reference genes, optionally extracts genomic coordinates, merges functional annotations, and generates publication-ready arrow plots. The package works seamlessly with or without the coding sequences input and skips plotting when no functional groups are found. For more details see Li et al. (2023) <[doi:10.1038/s41467-023-42193-7](https://doi.org/10.1038/s41467-023-42193-7)>.

URL <https://github.com/LiuyangLee/gclink>

BugReports <https://github.com/LiuyangLee/gclink/issues>

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Contents

blastp_df	2
eggnog_df	3
gclink	4
gc_add	8
gc_cal	9
gc_cluster	10
gc_plot	11
gc_position	12
gc_range	13
gc_scale	14
KO_group	15
length_filter	15
orf_extract	16
orf_locate	17
PGC_group	17
photosynthesis_gene_list	18
seq_data	18
Index	19

blastp_df	<i>BLASTP Results for test Proteins</i>
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Description

A dataset containing BLASTP alignment results for proteins from test genomes of bacteria, including alignment metrics.

Usage

blastp_df

Format

A data frame with multiple rows and 12 variables:

qaccver Character. Protein query identifier.

saccver Character. Subject protein identifier from reference databases.

pident Numeric. Percentage identity between query and subject sequences.

length Numeric. Length of the aligned sequence region.

mismatch Numeric. Number of mismatches in the alignment.

gapopen Numeric. Number of gap openings in the alignment.

qstart Numeric. Start position in query sequence.

qend Numeric. End position in query sequence.

sstart Numeric. Start position in subject sequence.
send Numeric. End position in subject sequence.
value Numeric. Expect value for the alignment.
bitscore Numeric. Bit score for the alignment.

 eggnog_df

EggNOG Functional Annotation Results

Description

A dataset containing EggNOG functional annotation results, including taxonomic classification, functional categories, and domain information.

Usage

eggnog_df

Format

A data frame with multiple rows and 21 variables:

#query Character. Protein query identifier (e.g., "Houyibacteriaceae-LLY-WYZ-15_3—k141_356661_13").

seed_ortholog Character. Subject protein identifier from EggNOG database (e.g., "1168065.DOK_14784").

value Numeric. Expect value for the annotation.

score Numeric. Annotation bitscore.

eggNOG_OGs Character.

max_annot_lvl Character.

COG_category Character.

Description Character. Functional description (e.g., "Bacterial regulatory proteins, tetR family").

Preferred_name Character. Gene name if available (e.g., "cobB").

GOs Character.

EC Character. Enzyme Commission number if applicable (e.g., "2.7.7.49").

KEGG_ko Character. KEGG Orthology identifier if available (e.g., "ko:K00986").

KEGG_Pathway Character.

KEGG_Module Character.

KEGG_Reaction Character.

KEGG_rclass Character.

BRITE Character.

KEGG_TC Character.

CAZy Character.

BiGG_Reaction Character.

PFAMs Character.

Description

`gclink()` performs **all steps** from raw `blastp` output to a publication-ready gene-cluster plot **in one call**. It dynamically adapts its workflow:

- If `in_seq_data` is provided: Extracts coordinates/sequences, merges data, and generates plots.
- If `in_seq_data` is `NULL`: Skips sequence-dependent steps, returning only the cluster table.
- If `in_GC_group` is `NULL`: Omits functional-group merging and plotting.
- Supports custom gene lists (e.g., photosynthesis, viral genes) via `in_gene_list` for universal cluster detection in bacteria, archaea, or phages.

Gene clusters are identified via a density threshold (`AllGeneNum` and `MinConSeq`) due to incomplete gene annotation coverage.

Usage

```
gclink(
  in_blastp_df = blastp_df,
  in_seq_data = seq_data,
  in_gene_list = photosynthesis_gene_list,
  in_GC_group = PGC_group,
  AllGeneNum = 50,
  MinConSeq = 25,
  apply_length_filter = FALSE,
  down_IQR = 10,
  up_IQR = 10,
  apply_evalue_filter = FALSE,
  min_evalue = 1,
  apply_score_filter = FALSE,
  min_score = 10,
  orf_before_first = 0,
  orf_after_last = 0,
  orf_range = "All",
  levels_gene_group = c("bch", "puh", "puf", "crt", "acsF", "assembly", "regulator",
    "hypothetical ORF"),
  color_theme = c("#3BAA51", "#6495ED", "#DD2421", "#EF9320", "#F8EB00", "#FF0683",
    "#956548", "grey"),
  genome_subset = NULL
)
```

Arguments

<code>in_blastp_df</code>	<p>A data.frame from blast/blastp-style output with columns:</p> <ul style="list-style-type: none"> • <code>qaccver</code>: Genome + contig name (separated by "---"), e.g., "Kuafubacteriaceae--GCA_016703535.1" <ul style="list-style-type: none"> – Genome: "Kuafubacteriaceae--GCA_016703535.1" ("--" separator), – Contig: "JADJBV010000001.1", – ORF: "JADJBV010000001.1_150" ("_" separator), – Position: "150". • <code>saccver</code>: Gene name + metadata (separated by "_"), e.g., "bchC_Methyloversatilis_sp_RAC08_B" <ul style="list-style-type: none"> – Gene: "bchC", – Metadata(Optional): "Methyloversatilis_sp_RAC08_BSY238_2447_METR". <p>eggNOG (evolutionary gene genealogy Nonsupervised Orthologous Groups) format results are supported by renaming annotation columns (e.g., "GOs") to <code>saccver</code>. Default: <code>blastp_df</code>.</p>
<code>in_seq_data</code>	<p>NULL or a data.frame with:</p> <p>SeqName ORF identifier (Prodigal format: >ORF_id # start # end # strand # ...). Sequence ORF sequence.</p> <p>Example: "Kuafubacteriaceae--GCA_016703535.1---JADJBV010000001.1_1 # 74 # 1018 # 1 # ..." Can be imported from Prodigal FASTA using:</p> <pre>seq_data <- Biostrings::readBStringSet("Prodigal.fasta", format="fasta", nrec=-1L, skip=1) data.frame(Sequence = .) %>% tibble::rownames_to_column("SeqName")</pre> <p>NULL skips coordinate extraction and plotting. Default: <code>seq_data</code>.</p>
<code>in_gene_list</code>	Character vector of reference genes for cluster detection. Default: <code>photosynthesis_gene_list</code> .
<code>in_GC_group</code>	NULL or a data.frame mapping genes to functional groups (columns: <code>gene</code> , <code>gene_group</code>). NULL skips plotting. Default: <code>PGC_group</code> .
<code>AllGeneNum</code>	Integer; max ORFs per cluster. Default: 50.
<code>MinConSeq</code>	Integer; min consecutive reference genes per cluster. Default: 25.
<code>apply_length_filter</code>	Logical; whether to apply length filtering based on IQR. If FALSE, skips <code>down_IQR</code> and <code>up_IQR</code> filtering. Default: FALSE.
<code>down_IQR</code>	Numeric; lower-bound scale factor for IQR length filtering (see <code>length_filter</code>). Default: 10.
<code>up_IQR</code>	Numeric; upper-bound scale factor for IQR length filtering (see <code>length_filter</code>). Default: 10.
<code>apply_evalue_filter</code>	Logical; whether to filter BLAST hits by e-value cutoff. Requires column <code>evalue</code> in input data. Default: FALSE.
<code>min_evalue</code>	Numeric; maximum e-value threshold for retaining BLAST hits. Hits with e-value > cutoff will be removed. Typical values: 1e-3 (loose) to 1e-10 (strict). Default: 1.

<code>apply_score_filter</code>	Logical; whether to filter BLAST hits by bitscore cutoff. Requires column <code>bitscore</code> in input data. Default: <code>FALSE</code> .
<code>min_score</code>	Numeric; minimum bitscore threshold for retaining BLAST hits. Hits with <code>bitscore < cutoff</code> will be removed. For short proteins (~100 aa), 30-40 is typical; for long proteins (>300 aa), 50-100 is recommended. Default: 10.
<code>orf_before_first</code>	Integer; number of hypothetical ORFs to insert before the first gene of each detected cluster. Useful when the upstream annotation is incomplete or low-confidence; insertion is bounded by the first ORF of the contig. Default: 0.
<code>orf_after_last</code>	Integer; number of hypothetical ORFs to append after the last gene of each detected cluster. Helpful when the downstream annotation is incomplete or low-confidence; insertion is bounded by the last ORF of the contig. Default: 0.
<code>orf_range</code>	Character. ORF inclusion mode: <ul style="list-style-type: none"> • "All": Include all ORFs + annotations (default) • "OnlyAnnotated": Keep only annotated ORFs • "IgnoreAnnotated": Include all ORFs without annotation merging
<code>levels_gene_group</code>	Character vector; factor levels for gene-group legends (must include "hypothetical ORF" in case some genes remain unclassified). Ignored if <code>in_GC_group</code> is <code>NULL</code> . Default: <code>c('bch', 'puh', 'puf', 'crt', 'acsF', 'assembly', 'regulator', 'hypothetical ORF')</code> .
<code>color_theme</code>	Character vector; colours for <code>gc_plot</code> , matched to the length and order of <code>levels_gene_group</code> . Ignored if plotting is skipped. Default: <code>c('#3BAA51', '#6495ED', '#DD2421', '#EF9320', '#F8EB00', '#F8EB00')</code> .
<code>genome_subset</code>	Character vector or <code>NULL</code> ; genomes to retain. If <code>NULL</code> , all genomes are retained. Default: <code>NULL</code> .

Value

A named list with:

`GC_meta` Annotated cluster table (data.frame).

`GC_seq` FASTA sequences (if `in_seq_data` provided).

`GC_plot` ggplot object (if `in_seq_data` and `in_GC_group` provided).

Examples

```
# Case 1: Using blastp result with Full pipeline (Find Cluster + Extract FASTA + Plot Cluster)
data(blastp_df)
data(seq_data)
data(photosynthesis_gene_list)
data(PGC_group)
gc_list <- gclink(in_blastp_df = blastp_df,
                 in_seq_data = seq_data,
                 in_gene_list = photosynthesis_gene_list,
                 in_GC_group = PGC_group,
                 AllGeneNum = 50,
```

```

        MinConSeq = 25,
        apply_length_filter = TRUE,
        down_IQR = 10,
        up_IQR = 10,
        orf_before_first = 0,
        orf_after_last = 0,
        levels_gene_group = c('bch', 'puh', 'puf', 'crt', 'acsF', 'assembly', 'regulator',
                              'hypothetical ORF'),
        color_theme = c('#3BAA51', '#6495ED', '#DD2421', '#EF9320', '#F8EB00',
                        '#FF0683', '#956548', 'grey'),
        genome_subset = NULL)
gc_meta = gc_list[["GC_meta"]]
gc_seq = gc_list[["GC_seq"]]
gc_plot = gc_list[["GC_plot"]]
head(gc_meta)
head(gc_seq)
print(gc_plot)

# Case 2: Using eggNOG result with Full pipeline (Find Cluster + Extract FASTA + Plot Cluster)
data(eggnog_df)
data(seq_data)
data(KO_group)
KOs = c("K02291", "K09844", "K20611", "K13789",
        "K09846", "K08926", "K08927", "K08928",
        "K08929", "K13991", "K04035", "K04039",
        "K11337", "K03404", "K11336", "K04040",
        "K03403", "K03405", "K04037", "K03428",
        "K04038", "K06049", "K10960", "K11333",
        "K11334", "K11335", "K08226", "K08226",
        "K09773")
rename_KOs = paste0("ko:", KOs)
eggnog_df$qaccver = eggnog_df$query`
eggnog_df$saccver = eggnog_df$KEGG_ko
eggnog_df$evalue = eggnog_df$evalue
eggnog_df$bitscore = eggnog_df$score
eggnog_df$gene = eggnog_df$KEGG_ko
gc_list_2 = gclink(in_blastp_df = eggnog_df,
                  in_seq_data = seq_data,
                  in_gene_list = rename_KOs,
                  in_GC_group = KO_group,
                  AllGeneNum = 50,
                  MinConSeq = 25,
                  apply_evalue_filter = FALSE,
                  min_evalue = 1,
                  apply_score_filter = TRUE,
                  min_score = 10,
                  orf_before_first = 1,
                  orf_after_last = 1,
                  levels_gene_group = c('bch', 'puh', 'puf', 'crt',
                                        'acsF', 'assembly', 'hypothetical ORF'),
                  color_theme = c('#3BAA51', '#6495ED', '#DD2421', '#EF9320',
                                   '#F8EB00', '#FF0683', 'grey'))
gc_meta_2 = gc_list_2[["GC_meta"]]

```

```

gc_seq_2 = gc_list_2[["GC_seq"]]
gc_plot_2 = gc_list_2[["GC_plot"]]
head(gc_meta_2)
head(gc_seq_2)
print(gc_plot_2)

# SOLUTION FOR "Viewport has zero dimension(s)" ERROR in print(gc_plot)
# -----
# When you see this error, try these 3 solutions:

# 1. RESIZE RSTUDIO PLOT PANEL (Quickest fix)
# Simply click and drag the right border of the plot panel wider

# 2. LAUNCH IN NEW WINDOW
# dev.new()
# print(gc_plot)

# 3. SAVE DIRECTLY TO FILE
# ggplot2::ggsave('gc_plot.pdf', gc_plot, width=20,height=3)

```

gc_add

Complete Gene Clusters by Adding Missing ORFs

Description

Expands gene cluster tables to include **all ORFs** (annotated and hypothetical) within contigs, normalizing cluster representations for downstream analysis and plotting. Ensures consistent ORF spacing/length across clusters by inserting missing rows.

Usage

```

gc_add(
  Data = sbgc,
  Annotation = bin_genes,
  orf_before_first = 0,
  orf_after_last = 0,
  orf_range = "All"
)

```

Arguments

Data	A data.frame of annotated ORFs with required columns: <ul style="list-style-type: none"> • qaccver: ORF identifier (genome—contig_orf_position format). • genome, contig: Genome and contig names. • gene: Gene symbol (NA for hypothetical ORFs). • orf_position: Absolute ORF position on the contig. • gene_cluster: Cluster identifier.
------	--

Annotation	A data.frame of full ORF annotations (e.g., from <code>orf_extract</code>). Must include <code>qaccver</code> and <code>orf_position</code> .
orf_before_first	Integer. Hypothetical ORFs to insert before the first annotated ORF in each cluster (bounded by contig start). Default: 0.
orf_after_last	Integer. Hypothetical ORFs to append after the last annotated ORF (bounded by contig end). Default: 0.
orf_range	Character. Controls ORF inclusion and annotation merging: <ul style="list-style-type: none"> • "All": Include every ORF in the contig range and merge all annotations (default). • "OnlyAnnotated": Keep only ORFs present in Annotation and merge their annotations. • "IgnoreAnnotated": Include all ORFs but skip merging with Annotation.

Details

- **Hypothetical ORFs** are inserted as rows with `gene = NA`.
- Output is always sorted by `gene_cluster` and `orf_position`.
- Progress messages are printed to console with timestamps.
- Contig bounds are respected—insertions never exceed actual ORF positions in Annotation.

Value

A data.frame with one row per ORF (real/hypothetical), sorted by `gene_cluster` and `orf_position`.
Added columns:

`GC_orf_position` Relative position within cluster (1-indexed).

`GC_present_length` Count of annotated ORFs in the cluster.

`GC_absent_length` Count of inserted hypothetical ORFs.

`GC_length` Total ORFs (`GC_present_length + GC_absent_length`).

gc_cal

Identify and Extract Gene Clusters from Scaled BLAST Data

Description

This function screens contigs for regions that contain a pre-defined set of “reference” genes (e.g., photosynthetic genes, viral genes) arranged in a continuous block. Contigs are first coarsely filtered by the minimum number of reference genes they carry, then finely scanned for clusters that satisfy user- defined density and contiguity criteria. Each detected cluster is returned with a unique `gene_cluster` identifier.

Usage

```
gc_cal(
  Data = bin_genes,
  in_gene_list = photosynthesis_gene_list,
  AllGeneNum = 30,
  MinConSeq = 15
)
```

Arguments

Data	A data frame produced by <code>orf_extract</code> (i.e., a scaled BLAST table). Must include the columns <code>genome_contig</code> , <code>gene</code> , and <code>orf_position</code> .
<code>in_gene_list</code>	A character vector of “reference” gene symbols (e.g., <code>photosynthesis_gene_list</code>) that are expected to appear in the target cluster(s).
AllGeneNum	Integer. Maximum total ORF count (annotated plus hypothetical) that the algorithm is allowed to span when defining a cluster (default: 30).
MinConSeq	Integer. Minimum number of reference genes that must be present and consecutive within the candidate cluster (default: 15). Must satisfy $1 \leq \text{MinConSeq} \leq \text{AllGeneNum}$.

Details

1. **Coarse filter:** Contigs with fewer than `MinConSeq` reference genes are discarded.
2. **Fine scan:** For each remaining contig, the algorithm slides a window that can encompass up to `AllGeneNum` consecutive ORFs and retains windows that contain at least `MinConSeq` reference genes in uninterrupted order.
3. **Cluster labelling:** Each valid cluster receives a unique ID (`genome_contig---1`, `genome_contig---2`, ...).

Value

A data frame identical in structure to `Data` but filtered to contain only those rows that belong to valid clusters. An extra column `gene_cluster` (format: `genome_contig---N`) is added to uniquely label every cluster.

`gc_cluster`

Identify Breakpoints of Gene Clusters within a Contig

Description

Internal helper used by `gc_cal`. Given the ordered positions of *reference* genes on a contig, this function returns the genomic coordinates that mark the boundary of each candidate cluster. A boundary is declared whenever the gap between two successive reference genes exceeds the maximum spacing allowed by the cluster definition (`AllGeneNum - MinConSeq`).

Usage

```
gc_cluster(Data = orf_position.tmp, AllGeneNum = 30, MinConSeq = 15)
```

Arguments

Data	A numeric vector (ascending order) of ORF positions that carry one of the reference genes of interest. Usually the vector <code>orf_position.tmp</code> created inside <code>gc_cal</code> .
AllGeneNum	Integer. Maximum genomic span (in ORF count) that the algorithm is allowed to cover when defining a single cluster. Passed unchanged from <code>gc_cal</code> .
MinConSeq	Integer. Minimum number of consecutive reference genes required to form a cluster. Passed unchanged from <code>gc_cal</code> .

Details

- If the gap between two consecutive reference genes is larger than `AllGeneNum - MinConSeq`, the left-hand gene is recorded as the **last member** of the preceding cluster.
- The final reference gene is always appended to the vector as the last boundary, ensuring the rightmost cluster is not lost.

Value

A numeric vector containing every position that marks the **end** of a putative gene cluster. These values are subsequently used as breakpoints to slice the contig into candidate regions in the downstream functions `gc_position()` and `gc_range()`.

gc_plot

Plot Scaled Gene Clusters with Arrows

Description

Generates a compact, publication-ready arrow plot of gene clusters previously processed with [gc_scale](#). Each cluster is displayed on its own horizontal track, with gene directionality, labels, and user-defined colour schemes.

Usage

```
gc_plot(
  data = GC_meta,
  color_theme = c("#3BAA51", "#6495ED", "#DD2421", "#EF9320", "#F8EB00", "#FF0683",
    "#956548", "grey")
)
```

Arguments

data	A data frame produced by gc_scale , must include the columns Pstart, Pend, Pgenome, gene_group, Pdirection, and gene_label.
color_theme	Character vector of colours, in the same order as the factor levels of gene_group. Defaults to an 8-colour palette; supply fewer or more colours as needed.

Value

A ggplot object that can be further customised or printed. The plot uses `gggenes::geom_gene_arrow()` and `gggenes::geom_gene_label()` with the following elements:

- One facet per Pgenome (cluster).
- Genes coloured by gene_group.
- Arrow direction encoded by Pdirection.
- Optional gene labels inside arrows.

gc_position

Extract ORF Positions for One Specific Gene Cluster

Description

Internal helper used by [gc_cal](#). Given the ordered positions of **all** reference genes on a contig (Data) and the vector of cluster breakpoints produced by [gc_cluster](#) (Cluster), this function slices Data to return the positions that belong to the EachCluster-th cluster.

Usage

```
gc_position(
  Data = orf_position.tmp,
  Cluster = cluster.tmp,
  EachCluster = eachcluster
)
```

Arguments

Data	A numeric vector (ascending order) of ORF positions that carry one of the reference genes of interest. Usually the vector <code>orf_position.tmp</code> created inside <code>gc_cal</code> .
Cluster	Numeric vector returned by gc_cluster ; each element marks the last position of a candidate cluster.
EachCluster	Integer index specifying which cluster to extract (1 = first cluster, 2 = second cluster, ...).

Value

A numeric vector containing the ORF positions that constitute the requested gene cluster.

`gc_range`*Determine ORF Range for a Candidate Gene Cluster*

Description

Internal helper used by `gc_cal`. After `gc_position` has isolated the ORF positions belonging to a single cluster, this function **validates** and **trims** that range so that the final span (distance between the first and last retained ORF) does not exceed `AllGeneNum`. The goal is to retain the **largest contiguous block** that still satisfies the user-defined size limit.

Usage

```
gc_range(Norf_position = Norf_position, AllGeneNum = 20, MinConSeq = 10)
```

Arguments

<code>Norf_position</code>	Numeric vector of ORF positions (ascending) that belong to the current candidate cluster (output from <code>gc_position</code>).
<code>AllGeneNum</code>	Integer. Maximum allowed genomic span (in ORF count) for the final cluster.
<code>MinConSeq</code>	Integer. Minimum number of consecutive reference genes required for the cluster.

Details

- For every reference gene in `Norf_position`, the function evaluates whether a window of at least `MinConSeq` consecutive reference genes centred on that gene can fit within `AllGeneNum` consecutive ORFs.
- Genes that pass the test are collected in `retain.site`.
- The **minimal** and **maximal** positions in `retain.site` are then used to slice the full ORF range, guaranteeing that the final cluster length \leq `AllGeneNum`.

Value

A numeric vector containing the **final ORF positions** that define the validated gene cluster. If no valid block can be produced, the vector will be empty.

gc_scale

*Scale Gene-Cluster Coordinates for Visualization***Description**

Prepares a gene-cluster annotation table for downstream plotting by converting absolute genomic coordinates into relative positions, ensuring that every cluster starts at 0 and is oriented consistently. Hypothetical ORFs (originally labeled with NA) and missing labels are replaced with placeholders, and factor levels are set as requested.

Usage

```
gc_scale(GC_meta = GC_meta, levels_gene_group = levels_gene_group)
```

Arguments

GC_meta A data frame containing gene-cluster information. Must include the columns `gene_cluster`, `gene_group`, `gene_label`, `start`, `end`, and `direction` (numeric: 1 for forward, -1 for reverse).

levels_gene_group Character vector specifying the desired factor levels for `gene_group`. Group names should appear in the order required for plotting legends.

Details

- Absolute start/end values are **not** modified; scaled values are stored in new columns (`Pstart`, `Pend`).
- `Pgenome` can be swapped for any unique identifier (e.g., `Genome`) downstream if each genome contains only one cluster.

Value

The input data frame with the following **new or overwritten** columns:

`gene_label` Empty string ("") if originally NA.

`gene_group` Set to "hypothetical ORF" if originally NA, then coerced to a factor using `levels_gene_group`.

`Pgenome` Factor version of `gene_cluster`; levels follow the order of appearance in the data.

`Pstart`, `Pend` Relative start and end coordinates (numeric) within each cluster, scaled so that the left-most gene starts at 0.

`Pdirection` Logical vector: TRUE for forward, FALSE for reverse.

KO_group	<i>KEGG Orthology (KO) Group Classification</i>
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Description

A dataset mapping KEGG Orthology (KO) identifiers to their functional groups and gene symbols.

Usage

KO_group

Format

A data frame with multiple rows and 3 variables:

gene Character. KEGG Orthology identifier (e.g., "K04035").

gene_group Character. Functional group (e.g., "acsF").

gene_label Character. Gene symbol (e.g., "acsF").

length_filter	<i>Remove Length Outliers from BLAST Results</i>
---------------	--

Description

Filters BLAST hits by removing ORFs whose gene (protein) length is an outlier within the corresponding gene group, as defined by the inter-quartile range (IQR). Hits whose length falls outside the interval $[Q1 - \text{down_IQR} * \text{IQR}, Q3 + \text{up_IQR} * \text{IQR}]$ are discarded.

Usage

```
length_filter(Data = bin_genes, down_IQR = 1.5, up_IQR = 1.5)
```

Arguments

Data	A data frame containing BLAST results. Must include the columns gene (gene symbol) and length (ORF length in amino acids).
down_IQR	Numeric multiplier applied to the IQR for the lower bound (default: 1.5).
up_IQR	Numeric multiplier applied to the IQR for the upper bound (default: 1.5).

Details

- Filtering is performed **within each gene group**; outliers are determined independently for every gene symbol.
- Progress messages report the number of rows before and after filtering.
- Missing values in length are ignored when computing quantiles.

Value

The input data frame with outlier rows removed. The returned object is **ungrouped** regardless of the input grouping.

orf_extract	<i>Extract ORF and Genome Information from BLAST or BLASTP Results</i>
-------------	--

Description

This function parses BLASTP result tables to extract structured genome, contig, ORF, and gene information from the query and subject identifiers. It is designed for downstream analyses requiring explicit separation of genome, contig, and ORF identifiers from concatenated BLAST headers.

Usage

```
orf_extract(bin_genes = blastp_df)
```

Arguments

bin_genes	A data frame containing BLASTP results with at least 2 standard columns: qaccver, saccver. the column of qaccver should include both of the genome name and predicted contig name, which is concatenated by a separator "—". for example, for the qaccver "p__Myxococcota—c__Kuafubacteria—o__Kuafubacteriales—f__Kuafubacteriaceae—GCA_016703535.1—JADJBV010000001.1_150", the genome name is "p__Myxococcota—c__Kuafubacteria—o__Kuafubacteriales—f__Kuafubacteriaceae—GCA_016703535.1", the contig name is "JADJBV010000001.1", the orf name is "JADJBV010000001.1_150", and the orf_position is "150". the column of saccver must include the gene name and may include the gene information, which are concatenated by a separator "_". for example, for the saccver "bchC_Methyloversatilis_sp_RAC08_BSY238_2447_METR", the gene name is "bchC", the gene information is Methyloversatilis_sp_RAC08_BSY238_2447_METR that can help you understand the source of gene.
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Value

The original data frame with six additional columns:

genome Genome identifier extracted from qaccver.

contig Contig identifier extracted from qaccver.

orf Full ORF identifier extracted from qaccver.

genome_contig Concatenated genome and contig IDs (genome—contig).

gene Gene symbol extracted from saccver.

orf_position Numeric ORF position extracted from the ORF identifier.

orf_locate *Parse ORF Coordinates from Prodigal FASTA Headers*

Description

Extracts ORF identifiers, start/end positions and strand orientation directly from the FASTA headers produced by Prodigal. The resulting table is ready for downstream gene-cluster analyses.

Usage

```
orf_locate(in_seq_data = seq_data)
```

Arguments

in_seq_data A data frame with two columns:
 SeqName ORF identifier (Prodigal format: >ORF_id # start # end # strand # ...).
 Sequence ORF sequence.

Example: "Kuafubacteriaceae--GCA_016703535.1---JADJBV010000001.1_1
 # 74 # 1018 # 1 # ..." Can be imported from **Prodigal** FASTA using:

```
seq_data <- Biostrings::readBStringSet("Prodigal.fasta", format="fasta", nrec=-1L, ski
data.frame(Sequence = .) %>%
  tibble::rownames_to_column("SeqName")
```

Value

A data frame

PGC_group *Photosynthesis Gene Classification Groups*

Description

A dataset mapping photosynthesis-related genes to their functional groups and subunits.

Usage

```
PGC_group
```

Format

A data frame with multiple rows and 3 variables:

gene Character. Gene identifier (e.g., "bchB").

gene_group Character. Functional group (e.g., "bch").

gene_label Character. Subunit designation (e.g., "B").

photosynthesis_gene_list

Photosynthesis Gene List

Description

A comprehensive list of genes involved in photosynthetic processes.

Usage

photosynthesis_gene_list

Format

A character vector containing gene identifiers:

Each element represents a photosynthesis-related gene symbol (e.g., "acsF", "bchB").

seq_data

Genomic Sequence Data with Annotations

Description

A dataset containing DNA sequences from test bacteria with detailed annotation metadata. The first column combines multiple annotation elements separated by semicolons.

Usage

seq_data

Format

A data frame with multiple rows and 2 variables:

SeqName Character. Combined annotation fields separated by semicolons, containing:

- ID: Sequence identifier (e.g., "1_7")
- partial: Completion status ("00" for complete, "01" for partial)
- start_type: Translation initiation codon (e.g., "GTG", "ATG")
- rbs_motif: Ribosome binding site motif (e.g., "GGAG/GAGG")
- rbs_spacer: RBS spacer length (e.g., "5-10bp")
- gc_cont: GC content (e.g., "0.673")

Sequence Character. DNA sequence (when available) in FASTA format

Index

* datasets

- blastp_df, [2](#)
- eggnoG_df, [3](#)
- KO_group, [15](#)
- PGC_group, [17](#)
- photosynthesis_gene_list, [18](#)
- seq_data, [18](#)

blastp_df, [2](#)

eggnoG_df, [3](#)

gc_add, [8](#)

gc_cal, [9](#), [10](#), [12](#), [13](#)

gc_cluster, [10](#), [12](#)

gc_plot, [11](#)

gc_position, [12](#), [13](#)

gc_range, [13](#)

gc_scale, [11](#), [12](#), [14](#)

gclink, [4](#)

KO_group, [15](#)

length_filter, [15](#)

orf_extract, [9](#), [10](#), [16](#)

orf_locate, [17](#)

PGC_group, [17](#)

photosynthesis_gene_list, [18](#)

seq_data, [18](#)