

Analysis of a GRTS Survey Design for a Finite Resource

Thomas Kincaid

May 23, 2011

Contents

1 Preliminaries	1
2 Read the survey design and analytical variables data file	1
3 Analysis of site status evaluation variables	3
4 Analysis of lake condition variables	7
5 Analysis of lake condition variables correcting for population size	8
6 Analysis of quantitative variables	10

This document presents analysis of a GRTS survey design for a finite resource. The finite resource used in the analysis is small lakes in Florida. The analysis will include calculation of three types of population estimates: (1) estimation of proportion and size (number of lakes) for site evaluation status categorical variables; (2) estimation of proportion and size for lake condition categorical variables; and (3) estimation of the cumulative distribution function (CDF) and percentiles for quantitative variables. Testing for difference between CDFs from subpopulations also will be presented.

1 Preliminaries

The initial step is to use the library function to load the `spsurvey` package. After the package is loaded, a message is printed to the R console indicating that the `spsurvey` package was loaded successfully.

Load the `spsurvey` package

```
> library(spsurvey)
```

Version 2.2 of the `spsurvey` package was loaded successfully.

2 Read the survey design and analytical variables data file

The original Florida small lakes data file contains more than 3,800 records and 29 basins. To produce a more manageable number of records, only six basins were retained in the data that will be analyzed, which produced a file containing 930 records.

The next step is to read the data file, which includes both survey design variables and analytical variables. The `read.delim` function is used to read the tab-delimited file and assign it to a data frame named `FL_lakes`. The `nrow` function is used to determine the number of rows in the `FL_lakes` data frame, and the resulting value is assigned to an object named `nr`. Finally, the initial six lines and the final six lines in the `FL_lakes` data frame are printed using the `head` and `tail` functions, respectively.

Read the survey design and analytical variables data file

```
> FL_lakes <- read.delim("FL_lakes.tab")
> nr <- nrow(FL_lakes)
```

Display the initial six lines in the data file.

```
> head(FL_lakes)
```

	siteID	xcoord	ycoord	wgt	basin	status	TNT
1	FLW03414-0014	8635535	12860896	5.369048	NFWWMD-1	Sampled	Target
2	FLW03414-0046	8636136	12886783	5.369048	NFWWMD-1	Physical Barrier	Target
3	FLW03414-0062	8617834	12869126	5.369048	NFWWMD-1	NonTarget	NonTarget
4	FLW03414-0078	8673500	12883071	5.369048	NFWWMD-1	Physical Barrier	Target
5	FLW03414-0086	8631884	12816428	5.369048	NFWWMD-1	NonTarget	NonTarget
6	FLW03414-0118	8607699	12856644	5.369048	NFWWMD-1	NonTarget	NonTarget

	pH_cat	coliform_cat	oxygen	turbidity
1	(0,6]	(0,5]	9.9	0.4
2	<NA>	<NA>	NA	NA
3	<NA>	<NA>	NA	NA
4	<NA>	<NA>	NA	NA
5	<NA>	<NA>	NA	NA
6	<NA>	<NA>	NA	NA

Display the final six lines in the data file.

```
> tail(FL_lakes)
```

	siteID	xcoord	ycoord	wgt	basin	status	TNT
925	FLW03414-3878	8880656	12694963	4.80791	SWFWMD-4	Dry	Target
926	FLW03414-3886	8892406	12732977	4.80791	SWFWMD-4	Sampled	Target
927	FLW03414-3894	8836528	12723056	4.80791	SWFWMD-4	Dry	Target
928	FLW03414-3918	8923107	12725502	4.80791	SWFWMD-4	Landowner Denial	Target
929	FLW03414-3926	8861298	12715824	4.80791	SWFWMD-4	Dry	Target
930	FLW03414-3950	8888601	12715641	4.80791	SWFWMD-4	NonTarget	NonTarget

	pH_cat	coliform_cat	oxygen	turbidity
925	<NA>	<NA>	NA	NA
926	(6,8]	(5,50]	1.98	8.2
927	<NA>	<NA>	NA	NA
928	<NA>	<NA>	NA	NA
929	<NA>	<NA>	NA	NA
930	<NA>	<NA>	NA	NA

The sample of small lakes in Florida is displayed in Figure 1. The sample sites for each basin are displayed using a unique color. First, the levels function is used to extract the set of basin names, and the result is assigned to object basin. Next, the rainbow function is called to select a set of six colors, and the result is assigned to object cols. The plot function is then used to produce the basic figure, but plotting of sample points is suppressed. The for function is used to loop through the set of six basins and plot color-coded points for each basin using the points function. Finally, the legend function is used to add a legend to the figure, and the title function is used to create a figure title.

```
> basins <- levels(FL_lakes$basin)
> cols <- rainbow(6)
> plot(FL_lakes$xcoord, FL_lakes$ycoord, type="n", xlab="x-coordinate",
+      ylab="y-coordinate")
> for(i in 1:6) {
+   ind <- FL_lakes$basin == basins[i]
+   points(FL_lakes$xcoord[ind], FL_lakes$ycoord[ind], pch=20, cex=0.4,
+         col=cols[i])
+ }
> legend(x="topright", inset=0.05, legend=basins, pch=20, cex=1, col=cols)
> title("Plot of Florida Small Lake Sites Color-Coded by Basin")
```

3 Analysis of site status evaluation variables

The first analysis that will be examined is calculation of extent estimates for site status evaluation variables. Extent is measured both by the proportion of the resource in status evaluation categories and by size of the resource in each category. For a finite resource like lakes, size refers to the number of lakes in a category. For calculating extent estimates (and for all of the analyses we will consider), the survey design weights are incorporated into the calculation process. Two site status variables will be examined: (1) status, which classifies lakes into six evaluation categories and (2) TNT, which classifies lakes as either "Target" or "NonTarget". The table and addmargins functions are used to create tables displaying the count for each code (level) of the two status variables.

```
> addmargins(table(FL_lakes$status))
```

A table displaying the number of values for each level of the status variable follows:

	Dry	Landowner Denial	NonTarget
	223	119	317
Otherwise Unsampleable		Physical Barrier	Sampled
	1	99	171
Sum			
	930		

```
> addmargins(table(FL_lakes$TNT))
```

A table displaying the number of values for each level of the TNT variable follows:

Plot of Florida Small Lake Sites Color-Coded by Basin

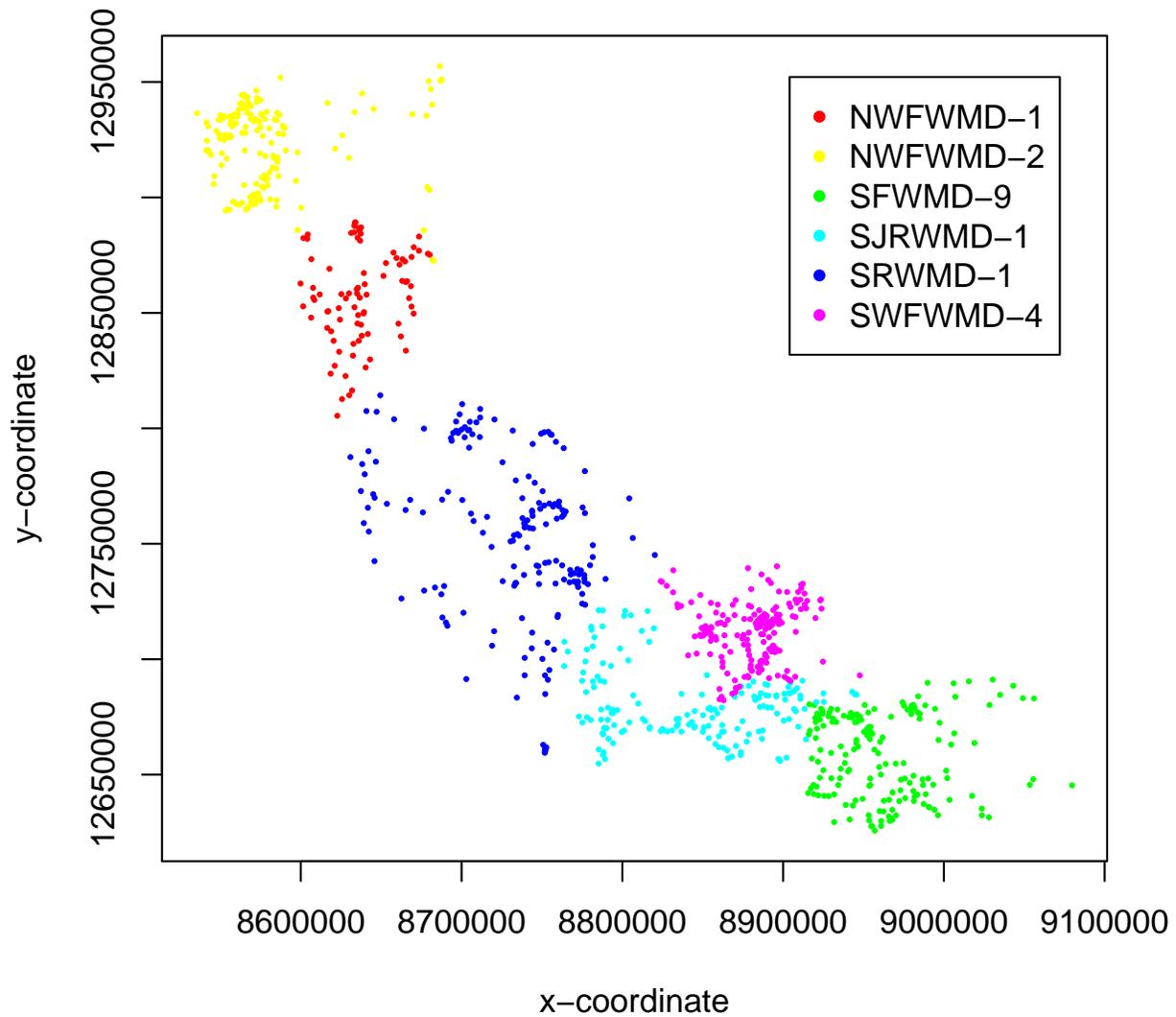


Figure 1: Florida Small Lake Sample Sites.

NonTarget	Target	Sum
317	613	930

The `cat.analysis` function in the `spsurvey` package will be used to calculate extent estimates. Four data frames constitute the primary input to the `cat.analysis` function. The first column (variable) in the four data frames provides the unique identifier (site ID) for each sample site and is used to connect records among the data frames. The `siteID` variable in the `FL_lakes` data frame is assigned to the `siteID` variable in the data frames. The four data frames that will be created are named as follows: `sites`, `subpop`, `design`, and `data.cat`. The `sites` data frame identifies sites to use in the analysis and contains two variables: (1) `siteID` - site ID values and (2) `Use` - a logical vector indicating which sites to use in the analysis. The `rep` (repeat) function is used to assign the value `TRUE` to each element of the `Use` variable. Recall that `nr` is an object containing the number of rows in the `FL_lakes` data frame. The `subpop` data frame defines populations and, optionally, subpopulations for which estimates are desired. Unlike the `sites` and `design` data frames, the `subpop` data frame can contain an arbitrary number of columns. The first variable in the `subpop` data frame identifies site ID values and each subsequent variable identifies a type of population, where the variable name is used to identify type. A type variable identifies each site with a character value. If the number of unique values for a type variable is greater than one, then the set of values represent subpopulations of that type. When a type variable consists of a single unique value, then the type does not contain subpopulations. For this analysis, the `subpop` data frame contains three variables: (1) `siteID` - site ID values, (2) `CombinedBasins` - which will be used to calculate estimates for all of the basins combined, and (3) `Basin` - which will be used to calculate estimates for each basin individually. The `basin` variable in the `FL_lakes` data frame is assigned to the `Basin` variable in the `subpop` data frame. The `design` data frame consists of survey design variables. For the analysis under consideration, the `design` data frame contains the following variables: (1) `siteID` - site ID values; (2) `wgt` - final, adjusted, survey design weights; (3) `xcoord` - x-coordinates for location; and (4) `ycoord` - y-coordinates for location. The `wgt`, `xcoord`, and `ycoord` variables in the `design` data frame are assigned values using variables with the same names in the `FL_lakes` data frame. Like the `subpop` data frame, the `data.cat` data frame can contain an arbitrary number of columns. The first variable in the `data.cat` data frame identifies site ID values and each subsequent variable identifies a response variable. The two response variables are `Status` and `Target_NonTarget`, which are assigned the `status` and `TNT` variables, respectively, in the `FL_lakes` data frame. Missing data (NA) is allowed for the response variables, which are the only variables in the input data frames for which NA values are allowed.

Create the `sites` data frame.

```
> sites <- data.frame(siteID=FL_lakes$siteID,
+                     Use=rep(TRUE, nr))
```

Create the `subpop` data frame.

```
> subpop <- data.frame(siteID=FL_lakes$siteID,
+                      CombinedBasins=rep("All Basins", nr),
+                      Basin=FL_lakes$basin)
```

Create the `design` data frame.

```
> design <- data.frame(siteID=FL_lakes$siteID,
+                      wgt=FL_lakes$wgt,
+                      xcoord=FL_lakes$xcoord,
+                      ycoord=FL_lakes$ycoord)
```

Create the data.cat data frame.

```
> data.cat <- data.frame(siteID=FL_lakes$siteID,
+                         Status=FL_lakes$status,
+                         Target_NonTarget=FL_lakes$TNT)
```

Use the cat.analysis function to calculate extent estimates for the site status evaluation variables.

```
> Extent_Estimates <- cat.analysis(sites, subpop, design, data.cat)
```

The extent estimates for all basins combined are displayed using the print function. The object produced by cat.analysis is a data frame containing thirteen columns. The first five columns identify the population (Type), subpopulation (Subpopulation), response variable (Indicator), levels of the response variable (Category), and number of values in a category (NResp). A category labeled "Total" is included for each combination of population, subpopulation, and response variable. The next four columns in cat.analysis provide results for the proportion estimates: the proportion estimate ((Estimate.P), standard error of the estimate (StdError.P), lower confidence bound (LCB95Pct.P), and upper confidence bound (UCB95Pct.P). Argument conf for cat.analysis allows control of the confidence bound level. The default value for conf is 95, hence the column names for confidence bounds contain the value 95. Supplying a different value to the conf argument will be reflected in the confidence bound names. Confidence bounds are obtained using the standard error and the Normal distribution multiplier corresponding to the confidence level. The final four columns in cat.analysis provide results for the size (units) estimates: the units estimate (Estimate.U), standard error of the estimate (StdError.U), lower confidence bound (LCB95Pct.U), and upper confidence bound (UCB95Pct.U).

```
> print(Extent_Estimates[c(1:7, 45:47), ])
```

	Type	Subpopulation	Indicator	Category	NResp					
1	CombinedBasins	All Basins	Status	Dry	223					
2	CombinedBasins	All Basins	Status	Landowner Denial	119					
3	CombinedBasins	All Basins	Status	NonTarget	317					
4	CombinedBasins	All Basins	Status	Otherwise Unsampleable	1					
5	CombinedBasins	All Basins	Status	Physical Barrier	99					
6	CombinedBasins	All Basins	Status	Sampled	171					
7	CombinedBasins	All Basins	Status	Total	930					
45	CombinedBasins	All Basins	Target_NonTarget	NonTarget	317					
46	CombinedBasins	All Basins	Target_NonTarget	Target	613					
47	CombinedBasins	All Basins	Target_NonTarget	Total	930					
			Estimate.P	StdError.P	LCB95Pct.P	UCB95Pct.P	Estimate.U	StdError.U		
1			23.01117939	0.97818958	21.093963	24.9283957	1184.155291	50.203673		
2			13.32737468	0.99082497	11.385393	15.2693559	685.826701	50.983555		
3			36.91250997	1.16116454	34.636669	39.1883507	1899.517763	60.323864		
4			0.09422536	0.08499027	0.000000	0.2608032	4.848837	4.373590		
5			8.47917794	0.71450383	7.078776	9.8795797	436.338497	36.735872		
6			18.17553265	1.03427992	16.148381	20.2026840	935.312910	53.208168		
7			100.00000000	0.00000000	100.000000	100.0000000	5146.000000	9.282499		
45			36.91250997	1.16116454	34.636669	39.1883507	1899.517763	60.323864		
46			63.08749003	1.16116454	60.811649	65.3633307	3246.482237	59.226647		

```

47 100.00000000 0.00000000 100.000000 100.0000000 5146.000000 9.282499
    LCB95Pct.U UCB95Pct.U
1   1085.7579 1282.55268
2    585.9008  785.75263
3   1781.2852 2017.75036
4     0.0000  13.42092
5    364.3375  508.33948
6    831.0268 1039.59900
7   5127.8066 5164.19336
45  1781.2852 2017.75036
46  3130.4001 3362.56433
47  5127.8066 5164.19336

```

The write.table function is used to store the extent estimates as a comma-separated value (csv) file. Files in csv format can be read by programs such as Microsoft Excel.

```

> write.table(Extent_Estimates, file="Extent_Estimates.csv", sep="," ,
+             row.names=FALSE)

```

4 Analysis of lake condition variables

The second analysis that will be examined is estimating resource proportion and size for lake condition variables. Two lake condition variables will be examined: (1) pH_cat, which classifies lakes by categories of pH value and (2) coliform_cat, which classifies lakes by categories of fecal coliform count. The table and addmargins functions are used to create tables displaying the count for each level of the two lake condition variables.

```

> addmargins(table(FL_lakes$pH_cat))

```

A table displaying the number of values for each level of the pH category variable follows:

(0,6]	(6,8]	(8,14]	Sum
78	82	11	171

```

> addmargins(table(FL_lakes$coliform_cat))

```

A table displaying the number of values for each level of the fecal coliform category variable follows:

(0,5]	(5,50]	(50,500]	(500,5e+03]	Sum
97	40	31	2	170

As for extent estimates, the cat.analysis function will be used to calculate condition estimates. The sites data frame for this analysis differs from the one used to calculate extent estimates. The Use logical variables in sites is set equal to the value "Sampled", so that only sampled sites are used in the analysis. The subpop and design data frames created in the prior analysis can be reused for this analysis. The data.cat data frame contains the two lake condition variables: pH_Cat and Coliform_Cat. Variables pH_cat and coliform_cat in the FL_lakes data frame are assigned to pH_Cat and Coliform_Cat, respectively.

Create the sites data frame.

```
> sites <- data.frame(siteID=FL_lakes$siteID,
+                       Use=FL_lakes$status == "Sampled")
```

Create the data.cat data frame.

```
> data.cat <- data.frame(siteID=FL_lakes$siteID,
+                         pHCat=FL_lakes$pH_cat,
+                         ColiformCat=FL_lakes$coliform_cat)
```

Use the cat.analysis function to calculate estimates for the lake condition variables.

```
> Condition_Estimates <- cat.analysis(sites, subpop, design, data.cat)
```

Print the lake condition estimates for all basins combined.

```
> print(Condition_Estimates[c(1:4, 28:32), ])
```

	Type	Subpopulation	Indicator	Category	NResp	Estimate.P		
1	CombinedBasins	All Basins	pHCat	(0,6]	78	42.915056		
2	CombinedBasins	All Basins	pHCat	(6,8]	82	50.396558		
3	CombinedBasins	All Basins	pHCat	(8,14]	11	6.688386		
4	CombinedBasins	All Basins	pHCat	Total	171	100.000000		
28	CombinedBasins	All Basins	ColiformCat	(0,5]	97	55.986933		
29	CombinedBasins	All Basins	ColiformCat	(5,50]	40	24.108155		
30	CombinedBasins	All Basins	ColiformCat	(50,500]	31	18.521502		
31	CombinedBasins	All Basins	ColiformCat	(500,5e+03]	2	1.383410		
32	CombinedBasins	All Basins	ColiformCat	Total	170	100.000000		
	StdError.P	LCB95Pct.P	UCB95Pct.P	Estimate.U	StdError.U	LCB95Pct.U	UCB95Pct.U	
1	2.8588764	37.311761	48.518350	401.39005	26.951025	348.56702	454.21309	
2	3.0201364	44.477200	56.315917	471.36552	28.641865	415.22849	527.50254	
3	1.5707294	3.609813	9.766959	62.55734	14.654986	33.83409	91.28058	
4	0.0000000	100.000000	100.000000	935.31291	7.420134	920.76971	949.85611	
28	2.8786468	50.344889	61.628977	519.19950	26.501223	467.25806	571.14095	
29	3.0407982	18.148300	30.068010	223.56900	28.550181	167.61167	279.52632	
30	2.4525384	13.714615	23.328389	171.76069	22.676735	127.31511	216.20628	
31	0.8259436	0.000000	3.002230	12.82917	7.665908	0.00000	27.85407	
32	0.0000000	100.000000	100.000000	927.35836	7.408810	912.83736	941.87937	

Use the write.table function to write the condition estimates as a csv file.

```
> write.table(Condition_Estimates, file="Condition_Estimates.csv", sep="," ,
+             row.names=FALSE)
```

5 Analysis of lake condition variables correcting for population size

The frame is a data structure containing spatial location data in addition to other attributes regarding a resource of interest and is used to create a survey design. A frame often takes the form of a shapefile. The frame can be used to obtain size values (e.g., number of lakes) for the

populations and subpopulations examined in an analysis. Examination of the Estimates.U column in the Condition_Estimates data frame produced by cat.analysis reveals that the estimated Total value for both condition variables and each combination of population value and subpopulation value does not sum to the corresponding frame size value. For example, the Total entry in the Estimate.U column for the pHcat variable, population "CombinedBasins" and subpopulation "All Basins" is 935 (rounded to a whole number). The corresponding frame size value is 5,146. The popsize (population size) argument to cat.analysis provides a mechanism for forcing the Total category to equal a desired value. First, the c (combine) function is used to create a named vector of frame size values for each basin. Output from the c function is assigned to an object named framesize. The popsize argument is a list, which is a particular type of R object. The popsize list must include an entry for each population type included in the subpop data frame, i.e., CombinedBasins and Basin for this analysis. The sum function applied to framesize is assigned to the CombinedBasins entry in the popsize list. Recall that the basin population type contains subpopulations, i.e., basins. When a population type contains subpopulations, the entry in the popsize list also is a list. The as.list function is applied to framesize, and the result is assigned to the Basin entry in the popsize list.

Assign frame size values.

```
> framesize <- c("NFWMD-1"=451, "NFWMD-2"=394, "SFWMD-9"=834, "SJRWMD-1"=1216,
+               "SRWMD-1"=1400, "SWFWMD-4"=851)
```

Use the cat.analysis function to calculate estimates for the lake condition variables.

```
> Condition_Estimates_popsiz <- cat.analysis(sites, subpop, design, data.cat,
+     popsize=list(CombinedBasins=sum(framesize),
+                 Basin=as.list(framesize)))
```

Print the lake condition estimates for all basins combined.

```
> print(Condition_Estimates_popsiz[c(1:4, 28:32), ])
```

	Type	Subpopulation	Indicator	Category	NResp	Estimate.P	StdError.P	LCB95Pct.P	UCB95Pct.P	Estimate.U	StdError.U	LCB95Pct.U	UCB95Pct.U
1	CombinedBasins	All Basins	pHcat	(0,6]	78	42.915056							
2	CombinedBasins	All Basins	pHcat	(6,8]	82	50.396558							
3	CombinedBasins	All Basins	pHcat	(8,14]	11	6.688386							
4	CombinedBasins	All Basins	pHcat	Total	171	100.000000							
28	CombinedBasins	All Basins	ColiformCat	(0,5]	97	55.986933							
29	CombinedBasins	All Basins	ColiformCat	(5,50]	40	24.108155							
30	CombinedBasins	All Basins	ColiformCat	(50,500]	31	18.521502							
31	CombinedBasins	All Basins	ColiformCat	(500,5e+03]	2	1.383410							
32	CombinedBasins	All Basins	ColiformCat	Total	170	100.000000							
1	2.8588764	37.311761	48.518350	2208.40876	147.11778	1920.0632	2496.7543						
2	3.0201364	44.477200	56.315917	2593.40689	155.41622	2288.7967	2898.0171						
3	1.5707294	3.609813	9.766959	344.18435	80.82973	185.7610	502.6077						
4	NA	NA	NA	5146.00000	NA	NA	NA						
28	2.8786468	50.344889	61.628977	2881.08756	148.13516	2590.7480	3171.4271						
29	3.0407982	18.148300	30.068010	1240.60567	156.47948	933.9115	1547.2998						
30	2.4525384	13.714615	23.328389	953.11648	126.20763	705.7541	1200.4789						
31	0.8259436	0.000000	3.002230	71.19029	42.50306	0.0000	154.4948						
32	NA	NA	NA	5146.00000	NA	NA	NA						

Use the write.table function to write the condition estimates as a csv file.

```
> write.table(Condition_Estimates_popsiz, file="Condition_Estimates_popsiz.csv",
+           sep=",", row.names=FALSE)
```

6 Analysis of quantitative variables

The third analysis that will be examined is estimating the CDF and percentiles for quantitative variables. Two quantitative variables will be examined: (1) oxygen - dissolved oxygen value and (2) turbidity - turbidity value. The summary function is used to summarize the data structure of the two quantitative variables.

```
> summary(FL_lakes$oxygen)
```

Summarize the data structure of the dissolved oxygen variable:

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.	NA's
0.830	4.880	6.870	6.468	8.310	12.480	759.000

```
> summary(FL_lakes$turbidity)
```

Summarize the data structure of the turbidity variable:

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.	NA's
0.150	1.100	1.700	8.055	3.800	400.000	759.000

The cont.analysis function will be used to calculate estimates for quantitative variables. Input to the cont.analysis function is the same as input for the cat.analysis function except that the data frame containing response variables is named cont.data rather than cat.data. The sites, subpop, and design data frames created in the analysis of lake condition variables can be reused for this analysis. The data.cont data frame contains the two quantitative variables: DissolvedOxygen and Turbidity. Variables oxygen and turbidity in the FL_lakes data frame are assigned to DissolvedOxygen and Turbidity, respectively. The popsize argument is included in the call to cont.analysis.

Create the data.cont data frame.

```
> data.cont <- data.frame(siteID=FL_lakes$siteID,
+                        DissolvedOxygen=FL_lakes$oxygen,
+                        Turbidity=FL_lakes$turbidity)
```

Use the cont.analysis function to calculate CDF and percentile estimates for the quantitative variables.

```
> CDF_Estimates <- cont.analysis(sites, subpop, design, data.cont,
+                               popsize=list(CombinedBasins=sum(framesize),
+                               Basin=as.list(framesize)))
```

The object produced by cont.analysis is a list containing two objects: (1) CDF, a data frame containing the CDF estimates and (2) Pct, a data frame containing percentile estimates plus estimates of population values for mean, variance, and standard deviation. Format for the CDF data frame is analogous to the data frame produced by cat.analysis. For the CDF data frame, however,

the fourth column is labeled Value and contains the value at which the CDF was evaluated. Unlike the data frames produced by the other analysis functions we have examined, the Pct data frame contains only nine columns since there is a single set of estimates rather than two sets of estimates. In addition, the fourth column is labeled Statistic and identifies either a percentile or the mean, variance, or standard deviation. Finally, since percentile estimates are obtained by inverting the CDF estimate, the percentile estimates do not have a standard error value associated with them.

Use the write.table function to write the CDF estimates as a csv file.

```
> write.table(CDF_Estimates$CDF, file="CDF_Estimates.csv", sep="," ,
+             row.names=FALSE)
```

The cont.cdfplot function in spsurvey can be used to produce a PDF file containing plots of the CDF estimates. The primary arguments to cont.cdfplot are a character string containing a name for the PDF file and the CDF data frame in the CDF_Estimates object. In addition, we make use of the logx argument to cont.cdfplot, which controls whether the CDF estimate is displayed using a logarithmic scale for the x-axis. The logx argument accepts two values: (1) "", do not use a logarithmic scale and (2) "x" - use a logarithmic scale. For this analysis, dissolved oxygen is displayed using the original response scale and turbidity is displayed using a logarithmic scale.

Produce a PDF file containing plots of the CDF estimates.

```
> cont.cdfplot("CDF_Estimates.pdf", CDF_Estimates$CDF, logx = c("",
+             "x"))
```

Print the percentile estimates for dissolved oxygen for all basins combined.

```
> print(CDF_Estimates$Pct[1:10, ])
```

	Type	Subpopulation	Indicator	Statistic	NResp	Estimate
1	CombinedBasins	All Basins	DissolvedOxygen	5Pct	8	1.578342
2	CombinedBasins	All Basins	DissolvedOxygen	10Pct	17	2.285793
3	CombinedBasins	All Basins	DissolvedOxygen	25Pct	42	4.624982
4	CombinedBasins	All Basins	DissolvedOxygen	50Pct	83	6.809475
5	CombinedBasins	All Basins	DissolvedOxygen	75Pct	129	8.333775
6	CombinedBasins	All Basins	DissolvedOxygen	90Pct	153	9.428672
7	CombinedBasins	All Basins	DissolvedOxygen	95Pct	163	9.996570
21	CombinedBasins	All Basins	DissolvedOxygen	Mean	171	6.477253
31	CombinedBasins	All Basins	DissolvedOxygen	Variance	171	6.442747
41	CombinedBasins	All Basins	DissolvedOxygen	Std. Deviation	171	2.538257
		StdError	LCB95Pct	UCB95Pct		
1		0.9547733	2.003741			
2		1.7532574	3.384512			
3		4.1087030	5.506430			
4		6.5622078	7.141814			
5		7.9710307	8.553521			
6		9.0221821	9.885101			
7		9.7547248	10.458964			
21	0.148923428773653	6.1853680	6.769137			
31	0.562255761580149	5.3407460	7.544748			
41	0.110756285008553	2.3211784	2.755335			

Use the write.table function to write the percentile estimates as a csv file.

```
> write.table(CDF_Estimates$Pct, file="Percentile_Estimates.csv", sep="," ,
+             row.names=FALSE)
```

The cont.cdfctest function in spsurvey can be used to test for statistical difference between the CDFs from subpopulations. For this analysis we will test for statistical difference between the CDFs from the six basins. The cont.cdfctest function will test all possible pairs of basins. Arguments to cont.cdfctest are the same as arguments to cont.analysis. Since we are interested only in testing among basins, the subpop data frame is subsetted to include only the siteID and Basin variables. Note that the popsize argument was modified from prior examples to include only the entry for Basin.

```
> CDF_Tests <- cont.cdfctest(sites, subpop[,c(1,3)], design, data.cont,
+   popsize=list(Basin=as.list(framesize)))
```

The print function is used to display results for dissolved oxygen of the statistical tests for difference between CDFs for basins. The object produced by cont.cdfctest is a data frame containing eight columns. The first column (Type) identifies the population. The second and third columns (Subpopulation_1 and Subpopulation_2) identify the subpopulations. The fourth column (Indicator) identifies the response variable. Column five contains values of the test statistic. Six test statistics are available, and the default statistic is an F-distribution version of the Wald statistic, which is identified in the data frame as "Wald-F". The default statistic is used in this analysis. For further information about the test statistics see the help file for the cdf.test function in spsurvey, which includes a reference for the test for differences in CDFs. Columns six and seven (Degrees_of_Freedom_1 and Degrees_of_Freedom_2) provide the numerator and denominator degrees of freedom for the Wald test. The final column (p_Value) provides the p-value for the test.

```
> print(CDF_Tests[1:15, ])
```

	Type	Subpopulation_1	Subpopulation_2	Indicator	Wald_F
1	Basin	NFWMD-1	NFWMD-2	DissolvedOxygen	3.1247258
2	Basin	NFWMD-1	SFWMD-9	DissolvedOxygen	4.4870771
3	Basin	NFWMD-1	SJRWMD-1	DissolvedOxygen	20.2999095
4	Basin	NFWMD-1	SRWMD-1	DissolvedOxygen	0.3056232
5	Basin	NFWMD-1	SWFWMD-4	DissolvedOxygen	10.6749729
6	Basin	NFWMD-2	SFWMD-9	DissolvedOxygen	2.6194412
7	Basin	NFWMD-2	SJRWMD-1	DissolvedOxygen	6.0724001
8	Basin	NFWMD-2	SRWMD-1	DissolvedOxygen	2.7890870
9	Basin	NFWMD-2	SWFWMD-4	DissolvedOxygen	3.8350098
10	Basin	SFWMD-9	SJRWMD-1	DissolvedOxygen	12.8288933
11	Basin	SFWMD-9	SRWMD-1	DissolvedOxygen	6.0791083
12	Basin	SFWMD-9	SWFWMD-4	DissolvedOxygen	14.0910554
13	Basin	SJRWMD-1	SRWMD-1	DissolvedOxygen	16.9152459
14	Basin	SJRWMD-1	SWFWMD-4	DissolvedOxygen	5.2459605
15	Basin	SRWMD-1	SWFWMD-4	DissolvedOxygen	6.3983641
	Degrees_of_Freedom_1	Degrees_of_Freedom_2			p_Value
1		2	55		5.183980e-02
2		2	57		1.549813e-02

3	2	57	2.203619e-07
4	2	54	7.379299e-01
5	2	51	1.340804e-04
6	2	55	8.191417e-02
7	2	55	4.141268e-03
8	2	52	7.069389e-02
9	2	49	2.835610e-02
10	2	57	2.510995e-05
11	2	54	4.157854e-03
12	2	51	1.342917e-05
13	2	54	1.977917e-06
14	2	51	8.475765e-03
15	2	48	3.440842e-03

Use the `write.table` function to write CDF test results as a csv file.

```
> write.table(CDF_Tests, file = "CDF_Tests.csv", sep = ",", row.names = FALSE)
```