

# Nonparametric maximum likelihood estimation for random effect models in R

Vignette to R package **npmlreg**

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## 1 Introduction

Nonparametric maximum likelihood (NPML) estimation is an attractive tool for the fitting of generalized linear models with random effects, which can be considered as a special case of generalized linear mixed models (GLMMs). One crucial advantage of the NPML approach is that the random effect distribution does not need to be specified a priori, whereas the huge body of literature on GLMMs restricts nearly exclusively on normally distributed random effects. Further, complicated integrations are avoided by approximating the marginal likelihood by a simple finite mixture, for which standard fitting algorithms based on EM exist and can be applied. NPML for generalized linear models with random effects was previously implemented by Aitkin & Francis (1995) in the GLIM4 language, which is however no longer widely used. The main functions of this package, `alldist` (for overdispersion) and `allvc` (for variance component models), are modified and extended versions of their homonymous counterparts in GLIM4, and have been translated to R originally by Ross Darnell.

In this handbook the concept of NPML estimation is briefly explained (Section 2) and a variety of data examples are given (Section 3), which illustrate the functionalities of `alldist` and `allvc`. The R package **npmlreg** is available for download at

<http://www.nuigalway.ie/mathsje/npml.html>.

*Key Words:* Varying coefficient models, random effect models, mixed models, mixture models, Gaussian Quadrature, EM algorithm, Two-level models, exponential family regression models.

## 2 Random effect modelling with exponential family mixtures

Assume there is given a set of explanatory vectors  $x_1, \dots, x_n$  and a set of observations  $y_1, \dots, y_n$  sampled from an exponential family distribution<sup>1</sup>  $f(y_i|\beta, \phi_i)$  with dispersion parameter  $\phi_i$ . In a generalized linear model, predictors and response are assumed to be related through a link function  $h$ ,

$$\mu_i \equiv E(y_i|\beta, \phi_i) = h(\eta_i) \equiv h(x_i'\beta),$$

and the variance  $Var(y_i|\beta, \phi_i) = \phi_i v(\mu_i)$  depends on a function  $v(\mu_i)$  which is entirely determined by the choice of the particular exponential family. However, often the actual variance in the data is larger than the variance according to this strict mean-variance

---

<sup>1</sup>In the present implementation of `alldist`, Gaussssian, Poisson, Binomial, and Gamma distributed response are supported

relationship. This effect is commonly called overdispersion. Reasons for overdispersion might be e.g. correlation in the data or important explanatory variables not included in the model. In order to account for additional unexplained variability of the individual observations, a random effect  $z_i$  with density  $g(z)$  is included into the linear predictor<sup>2</sup>

$$\eta_i = \beta' x_i + z_i.$$

The marginal likelihood can now be written as

$$L = \prod_{i=1}^n \int f(y_i | z_i, \beta, \phi_i) g(z_i) dz_i \quad (1)$$

and can be approximated by a finite mixture

$$\prod_{i=1}^n \left\{ \sum_{k=1}^K f(y_i | z_k, \beta, \phi_k) \pi_k \right\} \equiv \prod_{i=1}^n \left\{ \sum_{k=1}^K f_{ik} \right\},$$

where  $z_k$  are the mass points and  $\pi_k$  their masses. The log-likelihood is then given by

$$\ell = \sum_{i=1}^n \log \left\{ \sum_{k=1}^K \pi_k f_{ik} \right\}. \quad (2)$$

The score equations

$$\frac{\partial \ell}{\partial z_k} = 0, \quad \frac{\partial \ell}{\partial \beta} = 0, \quad \frac{\partial \ell}{\partial \phi_k} = 0, \quad (3)$$

turn out to be weighted versions of the single-distribution score equations, with weights

$$w_{ik} = \frac{\pi_k f_{ik}}{\sum_{\ell} \pi_{\ell} f_{i\ell}}. \quad (4)$$

---

<sup>2</sup>We refer to a model defined in this manner as a *generalized linear model with random effect*, or shorter, *random effect model*, whereas the more general linear predictor  $\eta_i = \beta' x_i + \gamma_i' \tilde{x}_i$ , with  $\gamma_i$  typically being a subvector of  $x_i$ , entails a *generalized linear mixed model*.

The weights  $w_{ik}$  can be interpreted as posterior probabilities that the observation  $y_i$  comes from component  $k$ . The score equation for the mixture proportions,

$$\frac{\partial \ell - \lambda (\sum \pi_k - 1)}{\partial \pi_k} = 0,$$

gives the ML estimate

$$\hat{\pi}_k = \frac{1}{n} \sum_i w_{ik} \tag{5}$$

which can be nicely interpreted as the average posterior probability for component  $k$ . The parameters  $\phi_k$ ,  $\beta$ ,  $z_k$  and  $\pi_k$  can now be simultaneously estimated by an standard EM algorithm:

**Starting points** Select starting values  $\phi^{(0)}$ ,  $\beta^{(0)}$ ,  $z_k^{(0)}$ , and  $\pi_k^{(0)}$ ,  $k = 1, \dots, K$ .

**E-Step** Adjust weights using formula (4) with current parameter estimates.

**M-Step** Update parameter estimates fitting a weighted GLM, including mass points as dummy variables.

In the special case of a normally distributed random effect, one can employ tabulated Gauss-Hermite integration points and their corresponding masses for  $z_k$  and  $\pi_k$ , respectively, and consider these values as constants (Hinde, 1982). Otherwise, they have to be calculated simultaneously during the EM algorithm as outlined above (one then usually takes the GH points/masses as starting points, which are scaled outwards `tol > 1` or inwards `tol < 1` by means of a scaling parameter `tol`). As in this case no parametric specification of the random effect distribution is necessary, one refers to this method as ‘Nonparametric Maximum Likelihood’ (NPML) estimation (Laird, 1978), which was adapted to the framework of overdispersed generalized linear models by Aitkin (1996a). In difference to the original implementation in GLIM4, we use a ‘damping’ procedure in the initial cycles of the algorithm, which reduces the sensitivity of the EM algorithm to the optimal choice of `tol` for exponential family densities possessing a dispersion parameter (as Gaussian or Gamma). For technical details on the implementation of the algorithm, see Einbeck & Hinde (2006).

## 3 Examples

### 3.1 Finite Gaussian mixtures: The galaxy data

The data considered in this example are the recession velocities (in  $km/s$ ) of 82 galaxies receding from our own, sampled from six well-separated conic sections of space. The full data were given by Postman et al. (1986). They are part of the R package **MASS** as data set *galaxies*. Note that, in this dataset, there is a typo in the 78th observation, which should be 26960 instead of 26690. We correct this to obtain consistent and comparable results with those presented in Aitkin et al. (2005) and other references.

```
> data(galaxies, package = "MASS")
> galaxies[78] <- 26960
> gal <- as.data.frame(galaxies)
> rm(galaxies)
```

Next, we construct a new variable `v1000` from `galaxies`, which represents the velocity in units of  $10^3 km/s$ :

```
> gal$v1000 <- gal$galaxies/1000
> gal$v1000
 [1]  9.172  9.350  9.483  9.558  9.775 10.227 10.406 16.084 16.170 18.419
[11] 18.552 18.600 18.927 19.052 19.070 19.330 19.343 19.349 19.440 19.473
[21] 19.529 19.541 19.547 19.663 19.846 19.856 19.863 19.914 19.918 19.973
[31] 19.989 20.166 20.175 20.179 20.196 20.215 20.221 20.415 20.629 20.795
[41] 20.821 20.846 20.875 20.986 21.137 21.492 21.701 21.814 21.921 21.960
[51] 22.185 22.209 22.242 22.249 22.314 22.374 22.495 22.746 22.747 22.888
[61] 22.914 23.206 23.241 23.263 23.484 23.538 23.542 23.666 23.706 23.711
[71] 24.129 24.285 24.289 24.366 24.717 24.990 25.633 26.960 26.995 32.065
[81] 32.789 34.279
```

and load the **npmlreg** package:

```

> library(npmlreg)
Lade nötiges Paket: statmod
      Fitting a simple constant normal model yields
> glm(v1000 ~ 1, data = gal)
Call:  glm(formula = v1000 ~ 1, data = gal)

Coefficients:
(Intercept)
20.83

Degrees of Freedom: 81 Total (i.e. Null); 81 Residual
Null Deviance:      1690
Residual Deviance: 1690    AIC: 484.8

which is the same as a NPML estimation with one mass point, fitting a 'mixture' of one normal component:
> (galaxy.np1 <- allidist(v1000 ~ 1, random = ~1, random.distribution = "np",
+   k = 1, data = gal))
Call:  allidist(formula = v1000 ~ 1, random = ~1, data = gal, k = 1,   random.distribution = "np")

Coefficients:
MASS1
20.83

MLE of sigma:      4.568
Mixture proportions:

```

```

MASS1
1
-2 log L: 480.8

The option data=... is mandatory, even if the data frame was attached to the workspace! The deviance can be obtained by
> galaxy.npi$dev
[1] 1690.296

which is certainly the same as for the GLM. Next, we fit discrete mixtures  $\sum_{k=1}^K \pi_k f_k$ , where the  $f_k$  are normal densities with expectation
 $\mu_k$  and unknown, but equal variances  $\sigma^2 = \sigma_k^2$ . Fitting models with  $K = 2, 3, 4$ , and 5 mass points, one obtains
> (galaxy.np2 <- alldist(v1000 ~ 1, random = ~1, random.distribution = "np",
+ k = 2, data = gal))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..19 ..20 ..21 ..22 ..23 ..24 ..25 ..26 ..27 ..28 ..29 ..30
EM algorithm met convergence criteria at iteration # 37
Disparity trend plotted.
EM Trajectories plotted.

Call: alldist(formula = v1000 ~ 1, random = ~1, data = gal, k = 2, random.distribution = "np")

Coefficients:
MASS1 MASS2
9.865 21.876

MLE of sigma: 3.026
Mixture proportions:
MASS1 MASS2
0.08694133 0.91305867
-2 log L: 461

```

```

> (galaxy.np3 <- allidist(v1000 ~ 1, random = ~1, random.distribution = "np",
+   k = 3, data = gal))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..
EM algorithm met convergence criteria at iteration # 10
Disparity trend plotted.
EM Trajectories plotted.

Call: allidist(formula = v1000 ~ 1, random = ~1, data = gal, k = 3,
  random.distribution = "np")

Coefficients:
MASS1 MASS2 MASS3
 9.75 21.40 32.94

MLE of sigma:      2.079
Mixture proportions:
  MASS1  MASS2  MASS3
0.08590000 0.87690389 0.03719611
-2 log L:      425.4

> (galaxy.np4 <- allidist(v1000 ~ 1, random = ~1, random.distribution = "np",
+   k = 4, data = gal))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..
EM algorithm met convergence criteria at iteration # 18
Disparity trend plotted.
EM Trajectories plotted.

Call: allidist(formula = v1000 ~ 1, random = ~1, data = gal, k = 4,
  random.distribution = "np")

```

```

Coefficients:
  MASS1  MASS2  MASS3  MASS4
  9.71  20.00  23.50  33.04

MLE of sigma:      1.315

Mixture proportions:
  MASS1  MASS2  MASS3  MASS4
  0.08536797  0.52624187  0.35180277  0.03658738
-2 log L:      416.5

```

and observes a steady decrease in disparity, i.e.  $-2 \log L$ . As a by-product, the `alldist` routine produces a plot showing how the disparity converges (Fig. 1 top), and another plot showing the EM trajectories (Fig. 1 bottom).

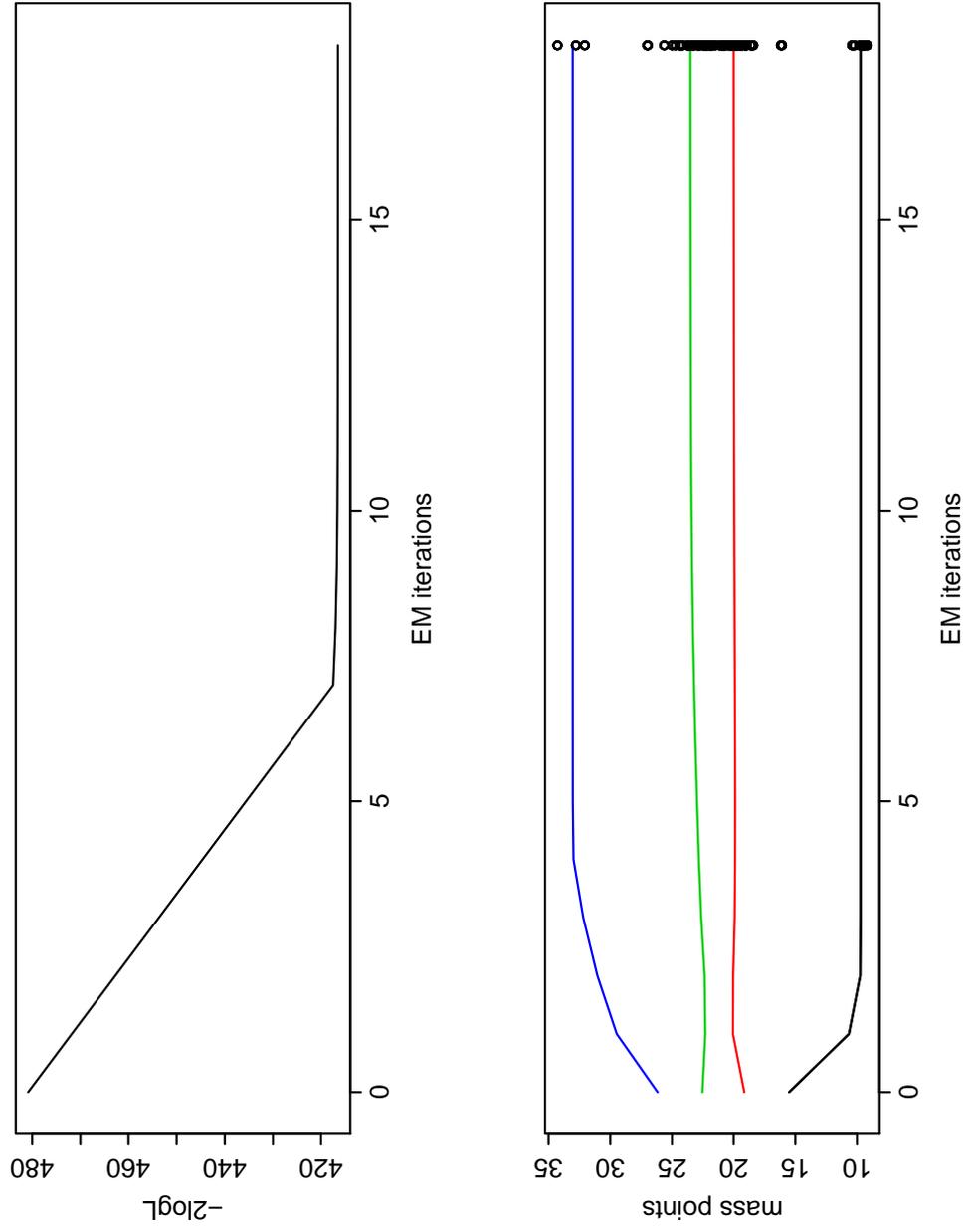


Fig. 1: Convergence of EM algorithm for NPML estimation with 4 mass points. Top: Disparities; Bottom: EM Trajectories.

As `random.distribution='np'` is the default setting, it can be omitted. For 5 to 9 mass points, we only report the disparity values

```
> (galaxy.np5 <- allidist(v1000 ~ 1, random = ~1, k = 5, data = gal,  
+ verbose = FALSE))$disp  
[1] 410.6852  
  
> (galaxy.np6 <- allidist(v1000 ~ 1, random = ~1, k = 6, tol = 0.2,  
+ data = gal, verbose = FALSE))$disp  
[1] 394.5811  
  
> (galaxy.np7 <- allidist(v1000 ~ 1, random = ~1, k = 7, tol = 0.12,  
+ data = gal, verbose = FALSE))$disp  
[1] 388.8639  
  
> (galaxy.np8 <- allidist(v1000 ~ 1, random = ~1, k = 8, tol = 0.2,  
+ data = gal, verbose = FALSE))$disp  
[1] 388.177  
  
> (galaxy.np9 <- allidist(v1000 ~ 1, random = ~1, k = 9, tol = 0.06,  
+ data = gal, verbose = FALSE))$disp  
[1] 388.2149
```

indicating that the disparity stabilizes at about 8 mass points. Note that in some cases it was necessary to modify the optional parameter `tol` to obtain the disparity values given above. The `tol` parameter influences the position of the starting points, where values `tol < 1` mean concentrated values compared to the default setting (Gaussian quadrature points). The disparity values for 2 and 5 mass points are better than those obtained by Aitkin (2001) with GLIM 4. One reason for that is the applied damping procedure: As the algorithm is

less sensitive to the optimal choice of `tol`, the optimal solutions are found more easily. An assisting tool in the selection of `tol` is the R function `tolfind` included in the package `npmlreg`.

To fit a Gaussian mixture with unequal standard deviations  $\sigma_k$ ,  $k = 1, \dots, K$  varying over the components, the possibility of smoothing the standard deviations among components is implemented. Smoothing is performed by means of the discrete kernel

$$W(x, y|\lambda) = \begin{cases} \lambda & \text{if } y = x \\ (1 - \lambda)/(K - 1) & \text{if } y \neq x \end{cases}$$

(Aitchison and Aitken, 1976). The setting  $\lambda = 1/K$  corresponds to the extreme case ‘maximal smoothing’ (i.e. equal variances  $\sigma^2 = \sigma_k^2$ ), while  $\lambda = 1$  means that all standard deviations are calculated within the components (i.e. unequal variances). Statistically sensible settings are only  $1/K \leq \lambda \leq 1$ . The default setting  $\lambda = 0$  is automatically mapped to  $\lambda = 1/K$ .

As an example, we compute the four mass-points model with option `lambda=1`

```
> summary(galaxy.np4u <- allidist(v1000 ~ 1, random = ~1, k = 4,
+   tol = 0.5, data = gal, lambda = 1, verbose = FALSE))
```

```
Call: allidist(formula = v1000 ~ 1, random = ~1, data = gal, k = 4,   tol = 0.5, lambda = 1, verbose = FALSE)
```

Coefficients:

	Estimate	Std. Error	t	value
MASS1	9.710143	0.2776679	34.97035	
MASS2	19.949549	0.1174379	169.87311	
MASS3	23.135282	0.1281410	180.54545	
MASS4	33.044336	0.4241453	77.90805	

Component standard deviations:

	MASS1	MASS2	MASS3	MASS4
0.4225107	1.3831150	1.6866727	0.9217176	

Mixture proportions:

```

      MASS1      MASS2      MASS3      MASS4
0.08536585  0.47707433  0.40097456  0.03658525
-2 log L:      405      Convergence at iteration 31

```

One gets deeper insight into the fitted model looking at diagnostic plots. Calling

```
> plot(galaxy.np4u, plot.opt = 15, height = 5)
```

gives the disparities and EM trajectories as above (Fig. 2 top), and additionally two plots showing the empirical Bayes predictions vs the true responses, and the component posterior probabilities ( $v_{ik}$ ) against the fixed part residuals ( $y_i - x_i\hat{\beta}$ ), respectively. In the former plot (Fig. 2 left bottom), one sees nicely how the predicted values are ‘flattened’ within the clusters and smoothed between. In the latter plot (Fig. 2 right bottom), one gets an impression of the discriminatory power of the mixture components. Throughout all plots, one colour corresponds to the same particular mass point.

An important remark should be given here: Interpretation (and definition!) of the deviance  $D = -2\phi \log L + 2\phi \log L_{saturated}$ , provided in **\$deviance**, is not clear when using unequal dispersion parameters. In the present implementation, deviances are calculated in this case in a somewhat makeshift manner using the *equal* dispersion parameter for the  $\phi$  preceding the log-likelihood, and the *unequal* dispersion parameters within the log-likelihood. Hence, it is strictly recommended to work with the disparity rather with the deviance in the case of unequal component dispersion parameters, as the the disparity does not share this problem. This is also the reason why we prefer to work with disparities in general, and why disparities (and not deviances) are displayed in the summaries.

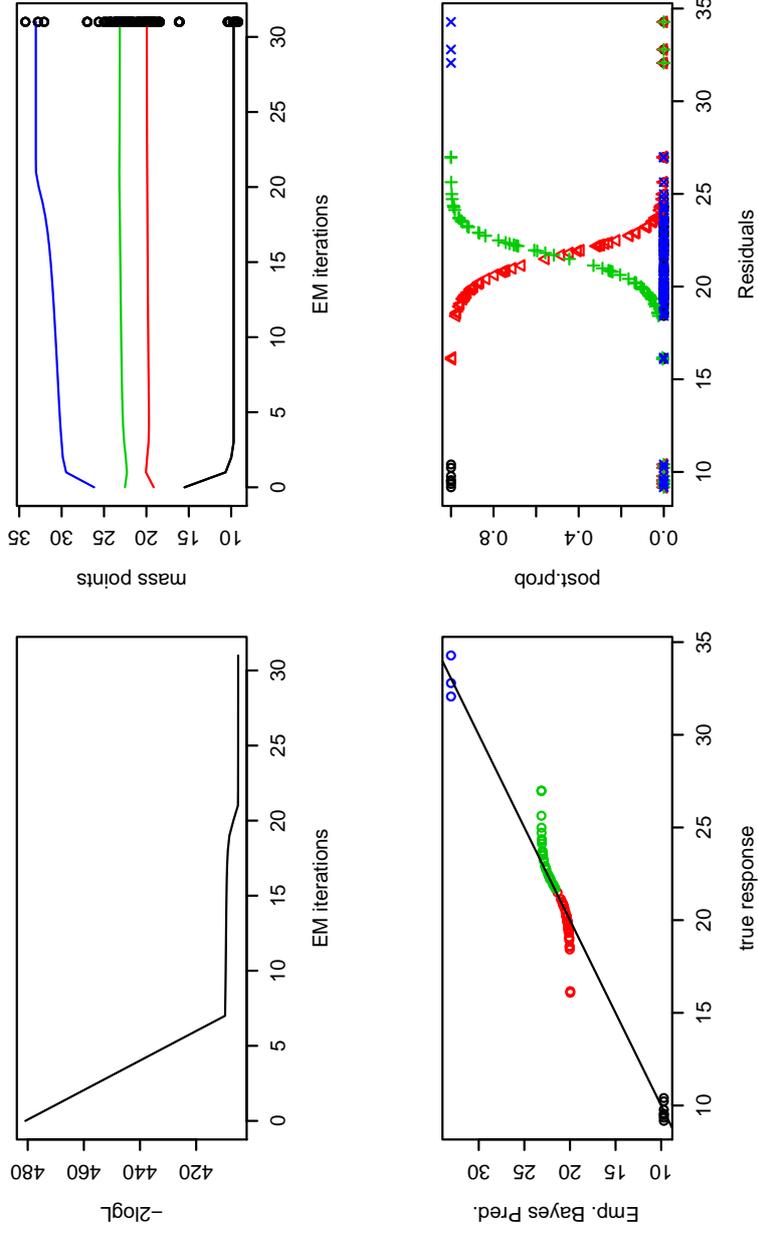


Fig. 2: Diagnostic plots for NPML estimation with unequal variances and 4 mass points.

One might fear that for a high number of mass points some component standard deviations could tend to zero. This can indeed be the case. Using 8 instead of 4 mass points in the call above one gets the error message

```
Error in glm.fit(x = XZ, y = Y, weights = as.vector(w) * pweights, family = family, :
  NAs in V(mu)
Error in alldist(v1000 1, random = 1, random.distribution = "np", k = 8, :
  Singularity or Likelihood-Spike at iteration #11. Enable spike protection or smooth among components.
```

This problem may be solved, as a first attempt, by modifying `tol`. In this case, `tol=0.32` gives a likelihood-spike free solution with disparity 357.8, which is a good part better than the value given in Aitkin (2001), 361.0. If likelihood spikes occur for any `tol`, one can enable the spike protection (`spike.protect=1`), which stops the algorithm as soon as one component starts to enter a likelihood spike. For instance, running

```
> (galaxy.mp8us <- alldist(v1000 ~ 1, random = ~1, k = 8, tol = 0.5,
+ data = gal, lambda = 1, verbose = FALSE, spike.protect = TRUE))
Call: alldist(formula = v1000 ~ 1, random = ~1, data = gal, k = 8, tol = 0.5, lambda = 1, spike.protect = TRUE, verbose = FALSE)

Coefficients:
  MASS1  MASS2  MASS3  MASS4  MASS5  MASS6  MASS7  MASS8
  9.383  9.906  17.233  19.772  22.587  24.410  32.433  34.279

Mixture proportions:
  MASS1  MASS2  MASS3  MASS4  MASS5  MASS6
  0.03457724  0.05078828  0.03637014  0.37478549  0.35365507  0.11323979
  MASS7  MASS8
  0.02438886  0.01219512
-2 log L: 220
> galaxy.mp8us$sdev$sdevk
```

```

[1] 1.504988e-01 4.113445e-01 1.526060e+00 6.309877e-01 1.170915e+00
[6] 1.738375e+00 3.757823e-01 2.645245e-37

```

gives us estimates of mass points, masses, and standard deviations of the mixture components. These values have to be interpreted with care, as the displayed disparity is normally not correct when the algorithm does not have converged. One notices from this output that the 8th mass point is responsible for the likelihood spike.

The better approach is to set the smoothing parameter equal to  $\lambda = 0.99$ , which corresponds to unequal standard deviations with a very low amount of smoothing among components:

```

> (galaxy.np8ud <- alldist(v1000 ~ 1, random = ~1, k = 8, tol = 0.5,
+ data = gal, lambda = 0.99))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..19 ..20 ..21 ..22 ..23 ..24 ..25 ..26 ..27 ..28 ..29 ..30
EM algorithm met convergence criteria at iteration # 102
Disparity trend plotted.
EM Trajectories plotted.

```

```

Call: alldist(formula = v1000 ~ 1, random = ~1, data = gal, k = 8, tol = 0.5, lambda = 0.99)

Coefficients:
MASS1 MASS2 MASS3 MASS4 MASS5 MASS6 MASS7 MASS8
9.836 9.710 16.127 19.790 22.922 26.978 32.427 34.279

Mixture proportions:
MASS1 MASS2 MASS3 MASS4 MASS5
7.88540e-15 8.536585e-02 2.439018e-02 4.039238e-01 4.256102e-01
MASS6 MASS7 MASS8
2.412464e-02 2.439092e-02 1.219444e-02
-2 log L:
374.6

```

```
> galaxy.mp8ud$sdev$sdevk
[1] 0.9061662 0.4349857 0.2183475 0.6758124 1.2048199 0.2160915 0.4119882
[8] 0.2949645
```

The motivation for the implementation of `spike.protect` is mainly to enable to run `tol.find` without breaking down if likelihood spikes occur. Hence, it is in `alldist` by default switched off, and in `tol.find` by default switched on. The result of `tol.find` for the 8-mass point model with unequal variances is shown in Fig. 2: Red circles correspond to `tol` values where the spike protection had to interfere and hence the EM algorithm did not converge. Only disparity values associated with green circles are reliable, and the optimal value of `tol` should consequently be chosen from them.

```

> par(mfrow = c(1, 1), cex = 0.65)
> tolfind(v1000 ~ 1, random = ~1, k = 8, data = gal, lambda = 1,
+ find.in.range = c(0, 0.6), steps = 12, plot.opt = 0, verbose = FALSE,
+ noformat = TRUE)[c(3, 4)]
Minimal Disparity: 219.9949 at tol= 0.5
Minimal Disparity with EM converged: 357.8216 at tol= 0.4
$AllDisparities
 [1] Inf 387.2360 377.6917 377.0345 360.9393 359.4521 357.8221 357.8216
 [9] Inf 219.9949 363.8772 Inf
$Alltol
 [1] 0.05 0.10 0.15 0.20 0.25 0.30 0.35 0.40 0.45 0.50 0.55 0.60

```

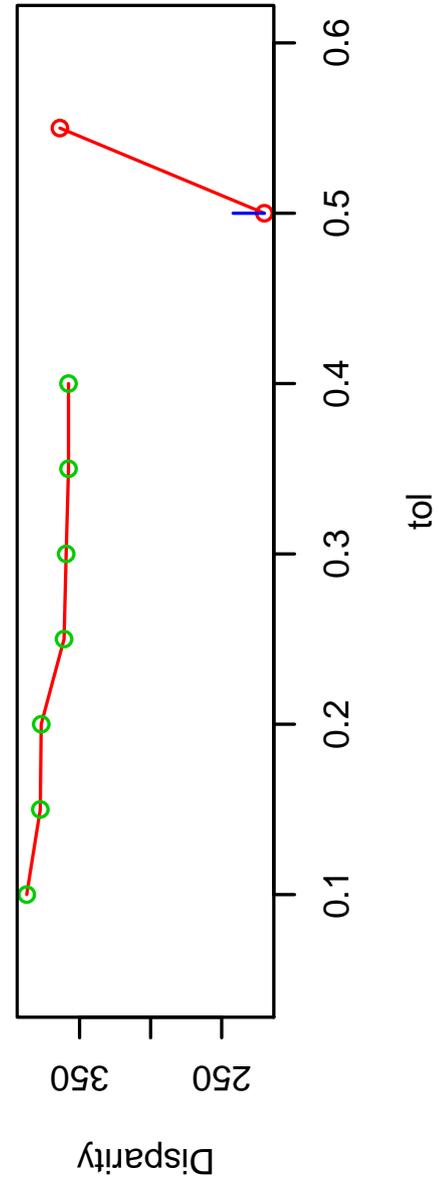


Fig. 3: Disparity against tol for 8 mass point model with unequal variances. Green circles: EM converged; red circles: EM not converged.

### 3.2 Compound Poisson models: The fabric faults data

In this Section we consider the fabric faults data, previously analyzed in Hinde (1982) and Aitkin, Francis & Hinde (2005, p. 453ff). This data describes the number of faults in rolls of fabrics with a single covariate `leng` for the length of the roll. The log-length is directly given by the variable `x`. The number of faults `y` can be assumed to follow a Poisson distribution. First, we fit a generalized linear Poisson model with the natural log link

```
> data(fabric, package = "gamlss")
> (faults0 <- glm(y ~ 1, family = poisson(link = log), data = fabric))
Call:  glm(formula = y ~ 1, family = poisson(link = log), data = fabric)
```

```
Coefficients:
(Intercept)
2.183
```

```
Degrees of Freedom: 31 Total (i.e. Null); 31 Residual
Null Deviance:
103.7
Residual Deviance: 103.7 AIC: 229
```

```
> (faults1 <- glm(y ~ x, family = poisson(link = log), data = fabric))
```

```
Call:  glm(formula = y ~ x, family = poisson(link = log), data = fabric)
```

```
Coefficients:
(Intercept)      x
-4.173         0.997
```

```
Degrees of Freedom: 31 Total (i.e. Null); 30 Residual
Null Deviance:
103.7
Residual Deviance: 64.54 AIC: 191.8
```

and observe a large reduction in deviance by including the log length. Fits of count data with Poisson models are often quite poor, as the basic assumption underlying a Poisson model, equality of mean and variance, is often not adequate. As a solution, Hinde (1982) proposed to model the unexplained variation by means of a Gaussian random effect  $Z$ . In case of the fabric fault data, one assumes that the number of faults conditional on the observation and on the random effect follows a Poisson distribution, i.e.

$$Y|X_1, \dots, X_n, Z \sim Po(\mu),$$

where  $Z \sim N(0, 1)$ , and

$$\log(\mu) = c + \log(\text{length}) + \sigma Z,$$

Integrating out the random effect as in (1), one obtains a Poisson/normal compound distribution, which can be approximated with Gaussian quadrature (GQ). For one, two and three mass points one obtains with the log length as covariate:

```
> (faults.g1 <- alldist(y ~ x, family = poisson(link = log), random = ~1,
+ data = fabric, k = 1, random.distribution = "gq"))
```

```
Call: alldist(formula = y ~ x, random = ~1, family = poisson(link = log), data = fabric, k = 1, random.distribution = "gq")
```

```
Coefficients:
```

```
  MASS1      x
-4.173  0.997
```

```
Mixture proportions:
```

```
  MASS1
      1      187.8
-2 log L:
```

```
> (faults.g2 <- alldist(y ~ x, family = poisson(link = log), random = ~1,
+ data = fabric, k = 2, random.distribution = "gq"))
```

```

1 ..2 ..3 ..4 ..5 ..6 ..
EM algorithm met convergence criteria at iteration # 6
Disparity trend plotted.

Call:  alldist(formula = y ~ x, random = ~1, family = poisson(link = log), data = fabric, k = 2, random.distribution = "gq")

Coefficients:
(Intercept)      x      z
-4.4128      1.0331      0.3391
-2 log L:
175.6

> (faults.g3 <- alldist(y ~ x, family = poisson(link = log), random = ~1,
+ data = fabric, k = 3, random.distribution = "gq", verbose = F))

Call:  alldist(formula = y ~ x, random = ~1, family = poisson(link = log), data = fabric, k = 3, random.distribution = "gq", verbose = F)

Coefficients:
(Intercept)      x      z
-3.3089      0.8488      0.3575
-2 log L:
174.3

The one mass point model is equivalent to the model faults1 given above, which can also be verified by checking the deviance

> faults.g1$dev
[1] 64.53719

```

For a Poisson model, deviance and disparity are related by the equation  $D = Disp + 2L_{sat}$  and are consequently equal up to an additive constant (the double saturated likelihood), which in our case takes the value 123.30. Thus, the disparity 174.3 for three mass points corresponds exactly to the deviance value of 51.0 reported in Hinde (1982), p. 119. For comparison, one can also fit the two and three mass point models with NPML:

```

> (faults.np2 <- alldist(y ~ x, family = poisson(link = log), random = ~1,
+ data = fabric, k = 2, random.distribution = "np"))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..19 ..20 ..21 ..22 ..
EM algorithm met convergence criteria at iteration # 22
Disparity trend plotted.
EM Trajectories plotted.

Call: alldist(formula = y ~ x, random = ~1, family = poisson(link = log), data = fabric, k = 2, random.distribution = "np")

Coefficients:
 x    MASS1    MASS2
0.8045 -3.1645 -2.4017

Mixture proportions:
MASS1    MASS2
0.7940023 0.2059977
-2 log L:      172.7

> (faults.np3 <- alldist(y ~ x, family = poisson(link = log), random = ~1,
+ data = fabric, k = 3, random.distribution = "np", verbose = F))
Call: alldist(formula = y ~ x, random = ~1, family = poisson(link = log), data = fabric, k = 3, random.distribution = "np", verbose =

Coefficients:
 x    MASS1    MASS2    MASS3
0.798 -3.154 -3.114 -2.353

Mixture proportions:
MASS1    MASS2    MASS3

```

```
0.1319235 0.6666708 0.2014057
-2 log L: 172.7
```

The disparities are not far from those obtained with Gaussian quadrature, indicating that the random effect distribution is not very far from a normal distribution. While three mass points seem to be adequate for GQ, only two mass points are needed with NPML. Note that the use of option `random.distribution="np"` yields an object of type `glimmNPML`, while option `random.distribution="gq"` yields an object of type `glimmGQ`.

Predictions for objects of type `glimmGQ` can be obtained by

```
> predict(faults.g2, type = "response", newdata = fabric[1:6, ])
```

```
1      2      3      4      5      6
8.7158 10.3546 13.3412 5.8568 11.4078 13.9380
```

or

```
> predict(faults.g2, type = "response")[1:6]
```

```
1      2      3      4      5      6
6.5578 7.0462 17.0202 7.2890 13.9926 9.5338
```

which both call function `predict.glimmGQ`. The results of the two predictions differ, since in the first case prediction is done using the analytical mean of the marginal distribution, considering `faults[1:6,]` as 'new' input data (Aitkin, Hinde & Francis, 2005, p. 459), and in the second case in an empirical Bayes approach (Aitkin, 1996a) using the individual posterior probabilities obtained as a by-product of the EM algorithm.

### 3.3 Logistic regression with random effects: The toxoplasmosis data

The toxoplasmosis data, also called rainfall data, are available via

```

> data(rainfall, package = "forward")
> rainfall$x <- rainfall$Rain/1000
> rainfall$x2 <- rainfall$x^2
> rainfall$x3 <- rainfall$x^3

```

gives the number of subjects Cases out of Total testing positively for toxoplasmosis in each of 34 cities in El Salvador with annual rainfall  $x$  in 1000 mm. The data have been analyzed in Efron (1998) using generalized linear models and in Aitkin & Francis (1995) using the GLIM 4 implementation of NPML. Fitting, as the latter authors, a constant logistic overdispersion model with three mass points, one obtains

```

> (toxop.np <- alldist(cbind(Cases, Total - Cases) ~ 1, random = ~1,
+ data = rainfall, k = 3, family = binomial(link = logit)))

```

```

1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..
EM algorithm met convergence criteria at iteration # 12
Disparity trend plotted.
EM Trajectories plotted.

```

```

Call: alldist(formula = cbind(Cases, Total - Cases) ~ 1, random = ~1, family = binomial(link = logit), data = rainfall, k = 3)

```

Coefficients:

```

MASS1 MASS2 MASS3
-0.9793 0.1492 0.7615

```

Mixture proportions:

```

MASS1 MASS2 MASS3
0.33421659 0.57062599 0.09515743
-2 log L:
146.9

```

The result is approximately the same as that obtained by Aitkin & Francis (1995). However, note that the disparity

```

> toxo.np$disparity
[1] 146.8668

```

differs from the GLIM 4 result 947.89. The disparity for binomial models provided by GLIM 4 has to be interpreted as  $-2 \log L + c$ , where  $c$  is some additive constant only depending on the values of  $y$  and  $n$ , while the disparity given by this R implementation is just  $-2 \log L$ . Adding rainfall as fixed effect, we fit a linear model

```

> (tox0.npx <- alldist(cbind(Cases, Total - Cases) ~ x, random = ~1,
+ data = rainfall, k = 3, family = binomial(link = logit)))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..
EM algorithm met convergence criteria at iteration # 12
Disparity trend plotted.
EM Trajectories plotted.

```

Call: `alldist(formula = cbind(Cases, Total - Cases) ~ x, random = ~1, family = binomial(link = logit), data = rainfall, k = 3)`

```

Coefficients:
 x      MASS1      MASS2      MASS3
0.2897 -1.5494 -0.4063  0.2385

```

Mixture proportions:

```

MASS1      MASS2      MASS3
0.3321544  0.5787877  0.0890579
-2 log L:
146.6

```

The decrease in disparity compared to the constant model is 0.3 on 1df (and is 5.1 for a cubic model, on 3df). We also try a random slope

```

> (tox0.npxx <- alldist(cbind(Cases, Total - Cases) ~ x, random = ~x,
+ data = rainfall, k = 3, family = binomial(link = logit)))

```

```

1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ...
EM algorithm met convergence criteria at iteration # 12
Disparity trend plotted.
EM Trajectories plotted.

Call: alldist(formula = cbind(Cases, Total - Cases) ~ x, random = ~x, family = binomial(link = logit), data = rainfall, k = 3)

```

```

Coefficients:
x MASS1 MASS2 MASS3 MASS1:x MASS2:x
-0.6688 -0.2451 -0.7948 2.0048 0.3038 1.1590

```

```

Mixture proportions:
MASS1 MASS2 MASS3
0.33569914 0.56715671 0.09714415
-2 log L:
146.1

```

giving only a negligible decrease in disparity compared to the linear fixed effects model. All in all, when accounting for overdispersion, there is no overwhelming evidence that rainfall has a significant influence on the incidence of toxoplasmosis at all.

For the simple constant model, the posteriori probabilities  $w_{ik}$

```

> round(t(toxo.np$post.prob), digits = 2)
  1  2  3  4  5  6  7  8  9  10 11 12 13 14 15
1 0.25 0.64 0.63 0.64 0.11 0.14 0.69 0.53 0.21 0.00 0.92 0.45 0.02 0.99 0.45
2 0.67 0.35 0.35 0.35 0.70 0.74 0.30 0.46 0.71 0.65 0.08 0.49 0.96 0.01 0.49
3 0.09 0.01 0.02 0.01 0.19 0.12 0.01 0.00 0.08 0.35 0.00 0.06 0.02 0.00 0.06
 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31
1 0.08 0.45 0.00 0.26 0.87 0.93 0.45 0.01 0 0.01 0.22 0.00 0.01 0.00 0.00 0.02
2 0.83 0.49 0.86 0.69 0.12 0.07 0.49 0.73 1 0.99 0.75 0.99 0.75 0.98 0.07 0.83
3 0.09 0.06 0.14 0.05 0.00 0.00 0.06 0.27 0 0.00 0.02 0.01 0.24 0.02 0.93 0.14

```

```

32 33 34
1 0.64 0.73 0.00
2 0.35 0.26 0.82
3 0.01 0.01 0.18

```

```
> plot(toxo.np, plot.opt = 3)
```

show the allocation of the data to the mass points, indicating that actually only one observation (the 30th) represents the 3rd mass point. From the posteriori probabilities, one also obtains the empirical Bayes predictions  $\tilde{\eta}_i = \sum_k \hat{\eta}_{ik} \hat{u}_{ik}$  as in Aitkin (1996b), from which the predicted toxoplasmosis incidence probabilities  $\tilde{p}_i = \exp(\tilde{\eta}_i) / (1 + \exp(\tilde{\eta}_i))$  can be calculated:

```
> round(toxo.ebp <- toxo.np$ebp, digits = 3)
```

```

1 2 3 4 5 6 7 8 9 10 11
-0.079 -0.566 -0.555 -0.566 0.146 0.065 -0.622 -0.450 -0.041 0.356 -0.885
12 13 14 15 16 17 18 19 20 21 22
-0.326 0.140 -0.971 -0.326 0.114 -0.326 0.237 -0.115 -0.838 -0.902 -0.326
23 24 25 26 27 28 29 30 31 32 33
0.304 0.152 0.139 -0.090 0.157 0.290 0.162 0.719 0.214 -0.566 -0.673
34
0.257

```

28

```
> round(exp(toxo.ebp)/(1 + exp(toxo.ebp)), digits = 4)
```

```

1 2 3 4 5 6 7 8 9 10 11
0.4803 0.3622 0.3647 0.3622 0.5363 0.5161 0.3494 0.3893 0.4899 0.5882 0.2921
12 13 14 15 16 17 18 19 20 21 22
0.4193 0.5350 0.2747 0.4193 0.5284 0.4193 0.5590 0.4714 0.3020 0.2886 0.4193
23 24 25 26 27 28 29 30 31 32 33
0.5754 0.5379 0.5347 0.4775 0.5393 0.5720 0.5403 0.6725 0.5534 0.3622 0.3378
34
0.5638

```

This can alternatively be done easier by using the generic predict function,

```
> predict(toxo.np, type = "response")
  1    2    3    4    5    6    7    8    9   10   11
0.4803 0.3622 0.3647 0.3622 0.5363 0.5161 0.3494 0.3893 0.4899 0.5882 0.2921
 12   13   14   15   16   17   18   19   20   21   22
0.4193 0.5350 0.2747 0.4193 0.5284 0.4193 0.5590 0.4714 0.3020 0.2886 0.4193
 23   24   25   26   27   28   29   30   31   32   33
0.5754 0.5379 0.5347 0.4775 0.5393 0.5720 0.5403 0.6725 0.5534 0.3622 0.3378
 34
0.5638
```

or, even quicker,

```
> fitted(toxo.np)
  1    2    3    4    5    6    7    8
0.4802916 0.3621653 0.3646799 0.3621653 0.5363293 0.5161203 0.3493630 0.3893228
  9   10   11   12   13   14   15   16
0.4898596 0.5881705 0.2920528 0.4193273 0.5350387 0.2747238 0.4193273 0.5283871
 17   18   19   20   21   22   23   24
0.4193273 0.5589992 0.4713726 0.3019893 0.2885689 0.4193273 0.5753992 0.5378508
 25   26   27   28   29   30   31   32
0.5347178 0.4775425 0.5392603 0.5719516 0.5403452 0.6724540 0.5533823 0.3621653
 33
0.3377802 0.5637801
```

which call function `predict.glmNPML` for an object of type `glmNPML`. The predict function can also be used to obtain predictions for new input values, e.g. for the linear model:

```
> predict(toxo.npx, type = "response", newdata = data.frame(x = 2))
1
0.4628
```

### 3.4 Modelling mixtures of Gamma distributions: The hospital stay data

The hospital-stay data is a sample from a larger data set collected on persons discharged from a Pennsylvania hospital as part of a retrospective chart review of antibiotic use in hospitals (Rosner, 2000, p. 39). Relevant covariates of the data set are `temp1` (the first measured temperature following admission, measured in Fahrenheit) and `age`, and the response is the duration of hospital stay. We read the data in and fit a three mass point model via

```
> data(hosp)
> (fitnp3 <- alldist(duration ~ age + temp1, data = hosp, k = 3,
+   family = Gamma(link = log), tol = 0.2))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..19 ..20 ..21 ..22 ..23 ..24 ..25 ..26 ..27 ...
EM algorithm met convergence criteria at iteration # 27
Disparity trend plotted.
EM Trajectories plotted.
```

```
Call: alldist(formula = duration ~ age + temp1, family = Gamma(link = log), data = hosp, k = 3, tol = 0.2)
```

```
Coefficients:
age          temp1          MASS1          MASS2          MASS3
0.004028  0.357537 -33.801628 -33.021757 -32.369541
```

```
MLE of shape parameter:          50.78
Mixture proportions:
MASS1      MASS2      MASS3
```

```

0.4798559 0.3980453 0.1220988
-2 log L:      121.3
giving the estimated shape parameter
> fitnp3$shape
$shape
[1] 50.78155

$shapek
[1] 50.78155 50.78155 50.78155

(Certainly, all three component shape parameters listed at $shape are equal, as by default all components are assumed to have the
same dispersion parameter.) For comparison, a three mass point mixture of exponentials (i.e. shape=1) is significantly inferior, yielding

> (fitnp3e <- alldist(duration ~ age + temp1, data = hosp, k = 3,
+   family = Gamma(link = log), tol = 0.2, shape = 1))
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..16 ..17 ..18 ..19 ..20 ..21 ..22 ..23 ..24 ..25 ..26 ..27 ..28 ..29 ..30
EM algorithm met convergence criteria at iteration # 41
Disparity trend plotted.
EM Trajectories plotted.

Call:  alldist(formula = duration ~ age + temp1, family = Gamma(link = log),      data = hosp, k = 3, tol = 0.2, shape = 1)

Coefficients:
age      temp1      MASS1      MASS2      MASS3
0.0149  0.3066 -28.6541 -28.6541 -28.6541

```

```

MLE of shape parameter:      1
Mixture proportions:
  MASS1      MASS2      MASS3
0.2295706  0.5465535  0.2238760
-2 log L:      155.2

```

For a more detailed analysis of this data set see Einbeck & Hinde (2006).

One important remark should still be given. When employing the damped EM algorithm, one will very occasionally observe rising disparities, typically from about the 10th to the 30th EM iteration. According to the theory on EM, this shouldn't happen. This phenomenon occurs as the damping step modifies the likelihood, and hence the theoretical basis for EM is not given any more. *Damped* EM is actually only *asymptotical* EM, as for a large number of iterations the effect of damping vanishes. It depends on the application and on `tol` what is a 'large' number of iterations. In general, rising disparities are more likely to be observed for small values of `tol` than for large values of `tol`, and more likely to be observed for Gamma than for Gaussian mixtures. Though these rising disparities are not really a problem, as at convergence the number of iterations is generally high enough, one might want to avoid them for esthetic reasons. This can be achieved by setting the constant `damp.power` in the formula

$$d_j = 1 - (1 - \text{tol})^{\text{damp.power} * \text{iter} + 1}$$

to a value bigger than one, where `iter` is the number of iterations and  $d_j$  is the multiplicative constant adjusting the dispersion parameter (Einbeck & Hinde, 2006). Another alternative is certainly to switch off damping (`option damp=FALSE`) but then it will not work at all in some cases.

### 3.5 Variance component models: The Oxford school boys data

This data set, also analyzed in Goldstein (2003), contains the heights of 26 boys in Oxford, measured on nine occasions over two years. The data set is contained in the R library `nlme` and can be loaded and plotted via

```

> data(Oxboys, package = "nlme")
> Oxboys$boy <- gl(26, 9)
> plot(Oxboys$age[Oxboys$boy == 1], Oxboys$height[Oxboys$boy ==
+ 1], ylim = c(125, 175), type = "b", pch = 1, xlab = "age",
+ ylab = "height")
> for (i in 2:nlevels(Oxboys$Subject)) {
+ lines(Oxboys$age[Oxboys$boy == i], Oxboys$height[Oxboys$boy ==
+ i], pch = 1, type = "b", col = i)
+ }

```

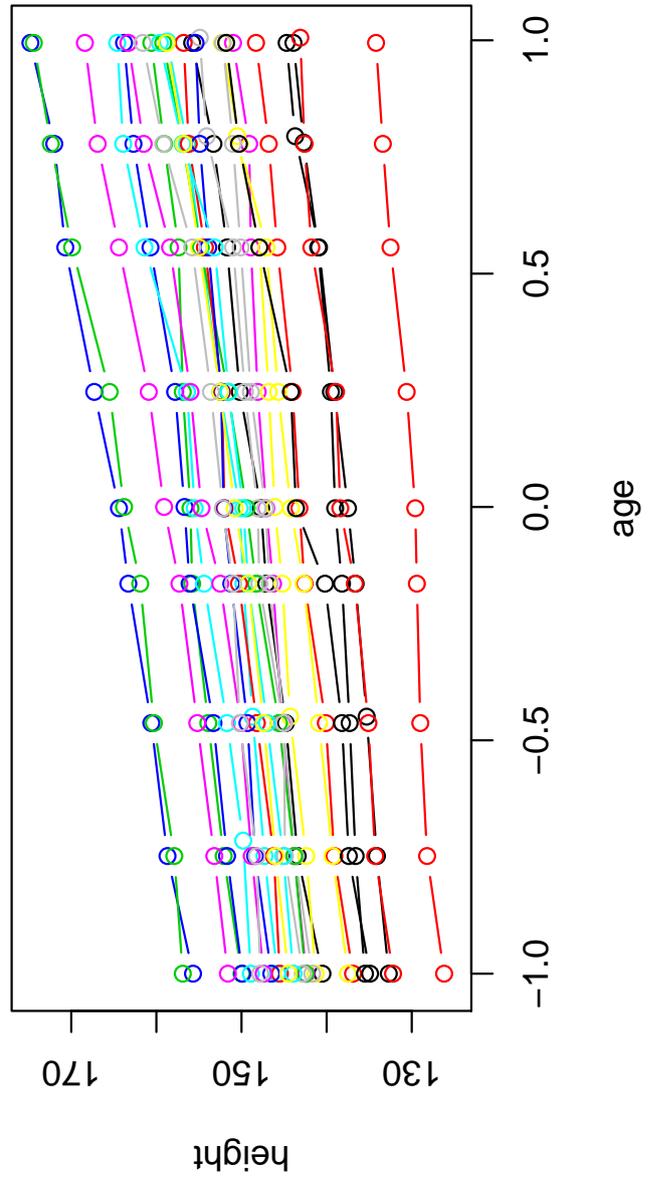


Fig. 4: Oxford Boys Data

The boys represent the upper level (primary sampling units, PSU), and the particular measurements at different time points correspond to the lower-level units (secondary sampling units, SSU). Fitting a variance component model with Gaussian quadrature (20 mass points), one gets

```
> (Oxboys.g20 <- allvc(height ~ age, random = ~1 | boy, data = Oxboys,
+   random.distribution = "gq", k = 20))
```

```
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..11 ..12 ..13 ..14 ..15 ..
```

```
EM algorithm met convergence criteria at iteration # 15
```

```
Disparity trend plotted.
```

```
Call: allvc(formula = height ~ age, random = ~1 | boy, data = Oxboys, k = 20, random.distribution = "gq")
```

```
Coefficients:
```

```
(Intercept)      age          z
148.958         6.524         4.769
```

```
MLE of sigma:      1.506
```

```
-2 log L:          991.8
```

For comparison, we fit NPML with seven (as suggested by Aitkin, Hinde & Francis (2005), p. 495), and eight masspoints, yielding

```
> (Oxboys.np7 <- allvc(height ~ age, random = ~1 | boy, data = Oxboys,
+   random.distribution = "np", k = 7))
```

```
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..
```

```
EM algorithm met convergence criteria at iteration # 10
```

```
Disparity trend plotted.
```

```
EM Trajectories plotted.
```

```
Call: allvc(formula = height ~ age, random = ~1 | boy, data = Oxboys, k = 7, random.distribution = "np")
```

```
Coefficients:
age MASS1 MASS2 MASS3 MASS4 MASS5 MASS6 MASS7
6.524 130.200 138.417 144.605 149.967 155.261 159.521 164.884
```

```
MLE of sigma: 1.762
```

```
Mixture proportions:
MASS1 MASS2 MASS3 MASS4 MASS5 MASS6 MASS7
0.03846154 0.11538462 0.19303307 0.34544795 0.19228146 0.03846828
```

```
0.07692308
-2 log L: 1017.3
```

```
33 > (Oxboys.np8 <- allvc(height ~ age, random = ~1 | boy, data = Oxboys,
+ random.distribution = "np", k = 8))
```

```
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..
EM algorithm met convergence criteria at iteration # 10
Disparity trend plotted.
EM Trajectories plotted.
```

```
Call: allvc(formula = height ~ age, random = ~1 | boy, data = Oxboys, k = 8, random.distribution = "np")
```

```
Coefficients:
age MASS1 MASS2 MASS3 MASS4 MASS5 MASS6 MASS7 MASS8
6.524 130.200 138.417 143.382 147.350 151.267 155.789 159.522
164.884
```

```

MLE of sigma:          1.433
Mixture proportions:
  MASS1      MASS2      MASS3      MASS4      MASS5      MASS6
0.03846154  0.11538462  0.11538469  0.19230765  0.26921962  0.15385725
  MASS7      MASS8
0.03846155  0.07692308
-2 log L:          931.4

```

Thus, NPMML with 8 mass points already leads to a better result than GQ with 20 mass points. The EM trajectories, as shown in Fig. 5, can also be obtained explicitly by calling

```
> plot(Oxboys.np8, plot.opt = 2)
```

We now extend the 8-point model by allowing the linear trend to vary across boys.

```
> (Oxboys.np8s <- allvc(height ~ age, random = ~age | boy, data = Oxboys,
+   random.distribution = "np", k = 8))
```

```
1 ..2 ..3 ..4 ..5 ..6 ..7 ..8 ..9 ..10 ..
```

EM algorithm met convergence criteria at iteration # 10

Disparity trend plotted.

EM Trajectories plotted.

```
Call: allvc(formula = height ~ age, random = ~age | boy, data = Oxboys, k = 8, random.distribution = "np")
```

```
Coefficients:
age      MASS1      MASS2      MASS3      MASS4      MASS5      MASS6
9.2130  130.2616  138.4476  143.3707  147.3756  151.2646  155.7763
MASS7      MASS8  MASS1:age  MASS2:age  MASS3:age  MASS4:age  MASS5:age
```

```

159.4738 164.8242 -5.4901 -4.0056 -2.1543 -3.7833 -2.5653
MASS6:age MASS7:age
-2.1239 -0.5421

```

```
MLE of sigma: 1.185
```

```
Mixture proportions:
```

```

MASS1 MASS2 MASS3 MASS4 MASS5 MASS6
0.03846154 0.11538462 0.11538462 0.19230769 0.26923047 0.15384645
MASS7 MASS8
0.03846154 0.07692308
-2 log L: 842.4

```

The difference in disparities is

```

> Oxbboys.np8$Disp - Oxbboys.np8s$Disp
numeric(0)

```

on 7df, showing clear heterogeneity in the slope.

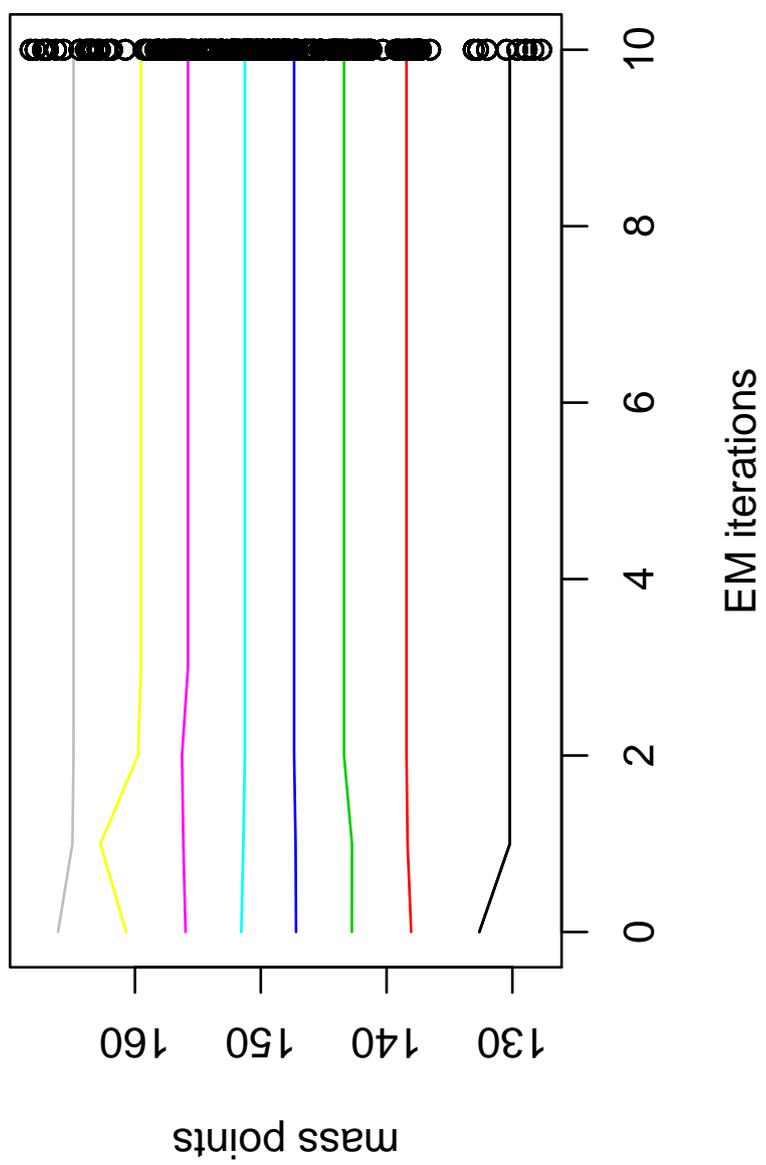


Fig. 5: Convergence of EM for eight mass points, applied on Oxford boys data.

### 3.6 Spatial random effect models: Irish Suicide Data

The data considered here, available in the package `npmlreg` via

```
> data(irlsuicide)
```

describe the mortality due to suicide and intentional self-harm in the Republic of Ireland from 1989–1998. Suicide rates are modelled using either the average crude rate or the relative risk as model parameter. The analysis of these data involves a variance component model with regions as cluster variable, categorical covariates for gender and age, interaction terms, a spatial autocorrelation term, and an offset. Thereby the spatial information for each region is included into the model by employing an extra - fixed or random - covariate representing the average crude suicide rates from the neighboring regions (or the average neighboring standard mortality ratios, respectively). For details, see Sofroniou, Einbeck and Hinde (2006) or the example in the helpfile for `allvc`.

## 4 Citation

The correct citation for R package `npmlreg` can be queried with

```
> citation(package = "npmlreg")
```

To cite package 'npmlreg' in publications use:

```
Jochen Einbeck, Ross Darnell and John Hinde (2006). npmlreg:  
Nonparametric maximum likelihood estimation for random effect models.  
R package version 0.38.
```

A BibTeX entry for LaTeX users is

```
@Manual{,  
  title = {npmlreg: Nonparametric maximum likelihood estimation for random effect  
models},
```

```
author = {Jochen Einbeck and Ross Darnell and John Hinde},
year = {2006},
note = {R package version 0.38},
}
```

ATTENTION: This citation information has been auto-generated from the package DESCRIPTION file and may need manual editing, see `help("citation")`.

The correct citation for this R vignette is

**EINBECK, J. and HINDE, J.** (2006). Nonparametric maximum likelihood estimation for random effect models in R. Vignette to R package ‘upmlreg’.

## Acknowledgments

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## 6 Appendix: R Documentation

The help files can be found at

<http://www.nuigalway.ie/maths/je/npmlreg/html/00Index.html>

or in the **npmlreg** Manual which can be downloaded in PDF format from

<http://www.nuigalway.ie/maths/je/npml.html> .

A list of all functions currently available in **npmlreg** is given below:

```
> ls("package:npmlreg")
[1] "alldist"          "allvc"          "binomial.expand"
[4] "dkern"           "expand"         "expand.vc"
[7] "family.glmnPML" "family.glmnPML" "gqz"
[10] "model.matrix.glmnPML" "model.matrix.glmnPML" "plot.glmnGQ"
[13] "plot.glmnPML"    "post"          "predict.glmnGQ"
[16] "predict.glmnPML" "print.glmnGQ"  "print.glmnPML"
[19] "summary.glmnGQ"  "summary.glmnPML" "tolfind"
[22] "weightslogl.calc.w"
```

In addition, the data sets `irlsucide`, `hosp`, and `missouri` are available.