

Simulating with Parameter Uncertainty

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1 Purpose

This script shows how to conduct a simulation that considers uncertainty in the parameter estimates.

2 Data

Here we load metrumrg and read in the data to be used for simulations.

Listing 1:

```
> library(metrumrg)
> data <- read.csv("../data/derived/phase1.csv")
> head(data)

      C ID TIME SEQ EVID   AMT     DV SUBJ HOUR TAFD    TAD LDOS MDV HEIGHT WEIGHT SEX
1 C  1 0.00  0    0   .     0   1 0.00 0.00   .   .   0   174  74.2   0
2 .  1 0.00  1    1 1000   .   1 0.00 0.00   0 1000  1   174  74.2   0
3 .  1 0.25  0    0   . 0.363   1 0.25 0.25 0.25 1000  0   174  74.2   0
4 .  1 0.50  0    0   . 0.914   1 0.50 0.50 0.5 1000  0   174  74.2   0
5 .  1 1.00  0    0   . 1.12   1 1.00 1.00   1 1000  0   174  74.2   0
6 .  1 2.00  0    0   . 2.28   1 2.00 2.00   2 1000  0   174  74.2   0
      AGE DOSE FED SMK DS CRCN predose zerodv
1 29.1 1000  1   0   0 83.5   1   0
2 29.1 1000  1   0   0 83.5   0   0
3 29.1 1000  1   0   0 83.5   0   0
4 29.1 1000  1   0   0 83.5   0   0
5 29.1 1000  1   0   0 83.5   0   0
6 29.1 1000  1   0   0 83.5   0   0
```

We use NONMEM output from a simple two compartment model to generate parameters. We use 1005.lst and 1005.csv output from NM7 to populate a call to metrumrg::simpar().

Listing 2:

```
> cov <- read.table("../nonmem/1005/1005.csv", skip=1, header=T)
> head(cov)

      NAME      THETA1      THETA2      THETA3      THETA4      THETA5
1 THETA1  0.85349000  0.78471700  1.02964e-03  0.06201550 -1.2885700
2 THETA2  0.78471700  4.74387000  6.65868e-03  0.89539600  5.5877600
3 THETA3  0.00102964  0.00665868  2.75169e-05  0.00221641 -0.0298637
4 THETA4  0.06201550  0.89539600  2.21641e-03  0.28656000  0.2410890
5 THETA5 -1.28857000  5.58776000 -2.98637e-02  0.24108900 559.0090000
6 THETA6 -0.03952260 -0.02453050 -1.02177e-04 -0.01047580  0.7350690
      THETA6      THETA7 SIGMA.1.1. SIGMA.2.1. SIGMA.2.2. OMEGA.1.1.
1 -0.03952260 -0.176224000 -4.89162e-04           0  2.04096e-02 6.28811e-03
2 -0.024530500 0.068529700 -3.11007e-03          0  1.89401e-02 5.84996e-03
3 -0.000102177 -0.000132916 -1.02493e-05          0  5.86438e-05 3.24081e-06
4 -0.010475800 0.015606300 -6.27671e-04          0  2.50369e-03 4.31368e-03
```

```

5  0.735069000 -0.684622000  4.52242e-02           0 -4.20659e-01 2.73881e-01
6  0.012748500  0.000415439  1.17741e-04           0 -1.03450e-03 1.63668e-03
    OMEGA.2.1.   OMEGA.2.2.   OMEGA.3.1.   OMEGA.3.2.   OMEGA.3.3.
1 -1.59957e-04 -4.31064e-03 -5.37918e-03 -2.56445e-03 -3.38999e-03
2 -2.19085e-02 -2.43988e-02 -1.95676e-02 -1.11920e-02  4.75058e-03
3 -6.49265e-05 -7.78059e-05 -6.74428e-05 -2.74920e-05  2.82116e-05
4 -6.19519e-03 -7.76509e-03 -4.54515e-03 -2.24113e-03  3.06880e-03
5  1.59962e-01  2.51679e-02 -7.08665e-03  7.40212e-02 -3.34805e-02
6  2.98890e-04  5.89470e-04 -5.36299e-04 -5.60638e-05 -3.30708e-04

```

We are interested in theta covariance, so we remove extra columns and rows.

Listing 3:

```
> cov<- cov[1:7, c(2:8)]
```

3 Parameters

Now we generate 10 sets of population parameters based on the 1005.lst results.

Listing 4:

```

> set.seed(10)
> PKparms <- simpar(
+   nsim=10,
+   theta=c(8.58,21.6, 0.0684, 3.78, 107, 0.999, 1.67),
+   covar=cov,
+   omega=list(0.196, 0.129, 0.107),
+   odf=c(40,40,40),
+   sigma=list(0.0671),
+   sdf=c(200)
+ )
> PKparms

```

	TH.1	TH.2	TH.3	TH.4	TH.5	TH.6	TH.7	OM1.1	OM2.2	OM3.3
1	9.458	24.04	0.06312	3.509	106.50	1.0150	1.593	0.1847	0.15400	0.13630
2	10.720	22.98	0.06798	3.817	111.30	0.8489	1.301	0.2862	0.12000	0.16400
3	9.024	21.24	0.06630	3.969	139.40	1.0770	1.507	0.1647	0.12770	0.11300
4	10.670	23.61	0.07183	3.909	121.20	1.1500	1.013	0.1886	0.11460	0.08460
5	10.010	23.01	0.07001	3.573	100.00	0.9469	1.767	0.1526	0.08448	0.13140
6	8.952	21.73	0.06876	3.343	97.78	1.0610	1.809	0.2462	0.17640	0.08805
7	8.655	19.73	0.06614	3.380	135.60	0.9608	1.343	0.2221	0.14440	0.09957
8	9.214	21.24	0.06098	3.084	115.60	1.0420	1.753	0.2287	0.13820	0.06118
9	8.998	23.97	0.06985	4.114	145.40	0.9439	1.778	0.1765	0.12310	0.08504
10	8.938	22.73	0.06436	3.704	113.10	1.0610	1.582	0.2116	0.11940	0.09954
	SG1.1									
1	0.06894									
2	0.06099									
3	0.06041									

```

4 0.07700
5 0.06269
6 0.07274
7 0.06160
8 0.06692
9 0.06092
10 0.06269

```

4 Control Streams

We read in a control stream and clean out extra xml markup.

Listing 5:

```

> ctl <- as.nmctl(readLines("../nonmem/ctl/1005.ctl"))
> ctl[] <- lapply(ctl,function(rec)sub("<.*","",rec))

```

Now we iterate across the rows of PKparms, writing out a separate ctl for each.

Listing 6:

```

> dir.create('../nonmem/sim')
> set <- lapply(
+     rownames(PKparms),
+     function(row,params,ctl) {
+         params <- as.character(PKparms[row,])
+         ctl$prob <- sub(1005,row,ctl$prob)
+         ctl$theta <- params[1:7]
+         ctl$omega <- params[8:10]
+         ctl$sigma <- params[11]
+         names(ctl)[names(ctl)=='estimation'] <- 'simulation'
+         ctl$simulation <- paste(
+             '(',
+             as.numeric(row) + 7995,
+             'NEW) (',
+             as.numeric(row) + 8996,
+             'UNIFORM) ONLYSIMULATION'
+         )
+         ctl$cov <- NULL
+         ctl$table <- NULL
+         ctl$table <- NULL
+         ctl$table <- 'ID TIME DV WT SEX LDOS NOPRINT NOAPPEND FILE=sim.tab
+
+         write.nmctl(ctl,file.path('../nonmem/sim',paste(sep='.',row,
+         ctl'))))
+         return(ctl)
+     },
+     params=PKparms,
+     ctl=ctl
+ )

```

5 Simulation

Finally, we run NONMEM simulations using NONR.

Listing 7:

```
> NONR72 ( 
+     run=1:10,
+     command="/opt/NONMEM/nm72/nmqual/autolog.pl",
+     project="../nonmem/sim",
+     diag=FALSE,
+     checkrunno=FALSE,
+     grid=TRUE
+ )
```